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1 **Uptake and distribution of ions reveal contrasting tolerance**
2 **mechanisms for soil and water salinity in okra (*Abelmoschus***
3 ***esculentus*) and tomatoes (*Solanum esculentum*)**

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10 Running title: Salinity tolerance mechanisms in tomato and okra

11 ***Abstract***

12 Okra and tomatoes are major vegetable crops commonly grown under irrigation, and
13 understanding whether they respond to salinity by withstanding (*tissue tolerance*) or avoiding
14 (*salt exclusion*) accumulation of salt in the shoots will assist with management for optimising
15 yield under declining soil and water resources. Both crops were grown in non-saline (0.0 dS/m)
16 and saline (3.0 dS/m) loamy sand and drip irrigated with water of 0.0, 1.2 or 2.4 dS/m.
17 Differences in the growth and yields of the two crops under saline conditions were associated
18 with uptake and distribution of cations, especially Na. The tomato employed *tissue tolerance*
19 mechanism in response to salinity and produced fruits even when shoot/root Na concentration
20 was >3.0; concentrations of Na in tomato tissues was in the order shoots > roots ≈ fruits. Okra

21 was sensitive to shoot Na such that a shoot/root Na concentration as low as 0.13 reduced yield by
22 as much as 35%; this crop thus employed *salt exclusion* mechanism and minimised shoot
23 accumulation of Na, which was distributed in the order fruits > roots > shoots. Root and shoot
24 concentrations of Na, P and S were correlated with flower abortion and negatively correlated
25 with yield and yield components in both crops. Fresh fruit produced on the saline soil were
26 reduced by 19% in tomato compared with 59% in okra, relative to yields on non-saline soil.
27 Water salinity reduced fresh fruit yields by as much as 36% with every unit (dS/m) rise in water
28 salinity compared with 27 % in okra. Soil salinity significantly reduced water-use by 6% in
29 tomatoes and 29% in okra, but had no impact on water use efficiency (WUE) that averaged 3.9 g
30 of fresh fruits/L for tomatoes and 1.75 g/L for okra. Every 1.0 dS/m rise in water salinity reduced
31 water-use by 0.33 L in okra and 3.31 L in tomatoes, and reduced WUE by 2.61 g/L in tomatoes
32 and 0.53 g/L in okra. Soil salinity explained <5% of the variance in yields in tomatoes and 10–
33 20% in okra, while water salinity explained 48–68 % of the variance in tomatoes and about 40%
34 in okra. We conclude that (1) water salinity was more injurious to yield in both crops than soil
35 salinity, and (2) yield losses due to salinity can be minimised through frequent leaching of soil
36 salt under okra and increased irrigation intervals in tomatoes.

37 **Keywords:** flower abortion, fruit yield, root growth, shoot/root Na, salinity, water-use, water-use
38 efficiency

39 **1.0 Introduction**

40

41 Crop species differ in their responses to saline conditions as a result of their relative tolerance to
42 ionic phytotoxicity. Two basic mechanisms that define crop tolerance of salinity involve ‘salt
43 exclusion’ or ‘tissue tolerance’, each of which is implemented to a varying degree by different

44 species with halophytes being adept almost equally at both (Munns et al, 2006). *Salt exclusion*
45 mechanism involves prevention of ions from getting into the transpiration stream by either
46 minimising their uptake from the growth media or if taken up expelling the ions into the
47 bathing/rooting medium, and/or restrained rates of root to shoot transfer. In *tissue tolerance*, on
48 the other hand, salts are sequestered in vacuoles of cells, especially in root tissue, thereby
49 restricting their transport into the cytoplasm of shoot tissues that are generally more sensitive to
50 salinity stress than roots, and where more physiological and enzymatic processes occur (Rogers
51 and West, 1993; Maas and Hoffman, 1977). Either or both of these mechanisms can be
52 overwhelmed resulting in phytotoxicity and death under extreme salinity.

53 Severity of impact of salinity on the plant also varies with the source of salinity, i.e. from water
54 or soil. Maas and Hoffman (1977) argued that plant response is primarily determined by the
55 salinity of the irrigation water rather than of the soil. This is because availability and uptake of
56 salt is governed by the availability of water and irrigation and/or rainfall reduces concentration of
57 salts especially in the top layer of soil where most plant roots reside; furthermore, the salts are
58 not available to the plant when the topsoil dries. They explained how salinity of the topsoil will
59 approximate that of the irrigation water, but will be more severe at the bottom of the root zone
60 (Maas and Hoffman, 1977). Such a situation should be particularly beneficial to plants that
61 exclude salts as the predominant mechanism for salinity tolerance.

62 Several ions have been associated with causing phytotoxicity under saline conditions and differ in
63 their adverse impact on plants (Shannon and Grieve, 1999). Amongst these, Na and Cl are the
64 most commonly associated with saline injury in plants, because they are easily accumulated in
65 shoot where they interfere with enzymatic, developmental and physiological processes (Flowers,
66 2004; Ghanem et al., 2009; Munns et al. 2006; Shannon and Grieve, 1999). Stunted plant growth

67 and reduced yields have often been associated with excessive Na and Cl concentrations in the
68 leaf that cause 'scorching' and 'firing' of leaves (Shannon and Grieve, 1999) and/or impairment
69 of CO₂ assimilation and photosynthetic capacity (Yunusa et al., 2009). Low yields, however,
70 could also result from late onset of reproductive phase and disruption of the processes involved.
71 In tomatoes, poor flower viability was associated with accumulation of Na at the expense of K in
72 the flower tissues and resulted in low fruit numbers, i.e. low sink capacity, and consequently
73 reductions in the overall fruit yield (Ghanem et al., 2009). Accumulation of Na in the leaves can
74 interfere with uptake of several other cations such as Ca, K and Mg. This can impair tolerance of
75 salinity, which is generally enhanced when plants selectively accumulate K relative to other
76 cations especially Na (Ashraf, 2004; Maksimović et al., 2010).

77 Tomato (*Lycopersicon esculentum* Mill.) and okra (*Abelmoschus esculentus* (L.) Moench) are
78 important vegetable crops commonly grown under irrigation. Extensive assessments of growth,
79 physiologic and biochemical responses to salinity have been undertaken for tomatoes (e.g.
80 Ghanem et al., 2009; Barbagallo et al., 2012; del Amor et al., 2001; Perez-Alfocea *et al.*, 2010),
81 but okra has received limited investigation in understanding its growth and yield responses to
82 ionic stress arising from media and/or water salinity. In this study, we compared ionic uptake
83 and partitioning, and their influence on the growth and yield of okra and tomatoes grown on
84 saline soil and irrigated with water of different salinities. The aims were to (1) quantify relative
85 tolerance to soil and water salinity, and (2) identify which of the two mechanisms of salinity
86 tolerance is dominant in the two crops.

87

88 **2.0 Materials and Methods**

89 **2.1 The Crops**

90 This study was undertaken in a glasshouse at the School of Environmental and Rural Sciences,
91 the University of New England, Armidale , Australia, over a 5-month period between March and
92 July in 2012. Tomato (*Solanum esculentum* 'Rouge de Marmande') and okra (*Abelmoschus*
93 *esculentus* 'Clemson's spineless') were raised from seeds obtained from a commercial supplier
94 (Mr Fothergill's Seeds of Australia[®]). The seeds were sown into vermiculite (0.0 dS/m) and
95 watered with tap water (EC of 0.025 dS/m) and they germinated within 6 days. The seedlings
96 were allowed to grow for 2 weeks (heights of 8–12 cm for okra and 10–18 cm for tomato),
97 before being transplanted into potted soils having different salinity. Three seedlings were
98 transplanted per pot then thinned down to two after 10 days and finally to one after 20 days.

99 **2.2 Salinity treatments**

100 A loamy sand soil (83% sand and 10% clay) having base salinity of 0.018 dS/m, pH of 6.27, and
101 water content at field capacity of 22% was collected from the nearby university research farm
102 (30° 29' 16" S, 151° 38' 29" E). Of this soil, 6 kg was weighed into each of 48 thick plastic bags.
103 Each bag was prepared to receive any one of the six treatments arising from factorial
104 combinations of the following:

- 105 • 2 levels of soil salinity: Control (0.018 dS/m) and saline (3.0 dS/m)
- 106 • 3 levels of water salinity: 0.025 dS/m (control), 1.2 dS/m (medium salinity) and 2.4 dS/m
107 (high salinity)

108 The soil salinity treatment of 3 dS/m was generated by adding 1% (w/w) table salt (NaCl) to half
109 the number of the bagged soil samples; the other half of the bagged soil samples received no salt.
110 The salinity and pH of the soil were determined using a bench top meter (Labchem-
111 CP[®]Benchtop Conductivity/TDS -pH/mV meter, TPS Pty Ltd., Brisbane, Australia).
112 All 48 bags received additional 2 kg soil that was pre-mixed with 2.5 g compound (12.2% N,
113 5.1% P, 13.7% K, 4.5% Ca and 1.1% Mg) fertiliser (Muriate of Potash, CSBP Ltd, Australia).
114 The bags were thoroughly shaken to achieve a homogeneous mixture. The bagged soil was then
115 transferred into separate, numbered plastic pots each having a diameter of 25 cm at the top, 19
116 cm at the base and a depth of 24 cm. The three levels of irrigation water salinity (denoted as 0,
117 1.2 and 2.4 dS/m) were obtained using tap water (EC, 0.025 dS/m) and dissolving 0, 88 or 225 g
118 NaCl/L, respectively. The tap water was considered as the control treatment. These solutions
119 were then stored in separate 220 L PVC tanks.

120 **2.3 Experimental layout and glasshouse weather**

121 The experimental units (pots) were laid out on benches in a glasshouse in a randomized design.
122 There were 24 pots per species, made up of two soil and three water salinity treatments in four
123 replicates. The glasshouse was maintained at a diurnal temperature range of 24–28°C and relative
124 humidity of 30–50%.

125 **2.4 Irrigation and nutrient management**

126 Each pot was supplied with a dripper that ran from a hose from the respective tank containing the
127 three saline solutions treatments. Each pot was irrigated at a rate of 100 mL for 5 min every day,
128 and was brought to field capacity every week to avoid water stress. Leachate was collected
129 separately from each pot every week and its volume determined. A 25 mL sub-sample of

130 leachate was stored in a dark cool room and later analysed for pH and EC, and the rest of the
131 leachate returned to their respective pots to maintain prescribed salinity for the pots. The salinity
132 of the water in the reservoirs was checked weekly to ensure that the prescribed salinity was
133 maintained.

134 All the pots were each supplied with 200 mL nutrient solution (16 g/L of Aquasol Hortico
135 containing NPK in 23:4:18) at 20 days after transplanting (DAT) and repeated when the plants in
136 the control treatments (non-saline soil and non-saline water) showed symptoms of nutrient
137 deficiency such as yellowing along the edges, curled leaves or early senescence of the older
138 leaves. Ten grams of dolomite (9% Mg and 14.5% Ca) was added to each pot to correct a Mg
139 deficiency for both crops evident by darkening of the fruit at the base in the control plants.

140 ***2.5 Measurements***

141 **2.5.1 Plant growth**

142 The height and leaf number for each plant was assessed every ten days, while leaf area was
143 determined on the last thinned plant at 20 DAT. Leaf area was measured with a scanning device
144 (CID Portable Leaf Area Meter CI-202, CID Bioscience Inc., Camas, WA, USA). The relative
145 chlorophyll concentration in the leaves was determined at 95 and 117 DAT using an optical
146 device (SPAD 502 Plus Chlorophyll Meter, Minolta, Japan); the SPAD readings were converted
147 to chlorophyll content according to Coste et al. (2010). Dates of appearance of first flower and
148 fruit were recorded, while numbers of flowers and fruits were counted daily. Flower abortion
149 was taken as the total number of fruits by the plant divided by the total number of flowers
150 counted for the same plant during its lifetime.

151 **2.5.2 Fruit yield and quality**

152 The fruits were carefully picked as they matured and weighed fresh. Weight of fruits harvested
153 from individual plants were collated and summed after picking the last fruit to determine total
154 yield. Sugar content of tomato fruit was determined on 1.0 ml squeezed juice using a hand-held
155 device (Cobras[®] Accutrend[®] Plus instrument, Roche Ltd, Schweiz, Switzerland).

156 The fruits along with the shoots were dried at 60° C for 72 h to determined dry weights. The
157 roots were recovered from the pots, thoroughly washed and also dried at 60° C for 72 h. Total dry
158 weight per plant was determined as the sum of dry weights of fruits, roots and shoots.

159 **2.5.3 Water use**

160 Amounts of water supplied to, and drained from, each pot was recorded and water-use was
161 obtained as: water-use (WU) = water applied (L) - water drained (L). The weekly values for WU
162 were summed at the end of the trial to obtain total amount of water used by the plant in each pot.
163 Water-use efficiency (WUE) was determined as: total weight of fresh fruit (kg)/WU (L).

164 **2.5.4 Elemental uptake and distribution**

165 Dried samples of the fruit, root and shoot tissues were ground separately using a mortar and
166 pestle to pass a 2 mm screen. Subsamples of the ground tissues (~0.5 g) were digested in
167 concentrated HNO₃ (70%) and H₂O₂ (30%) in a microwave digester. The digests were brought to
168 final volumes of 100 mL with double-deionized water, and the elemental contents determined
169 using ICP-MS (ICP-MS Agilent 7500CE, Agilent Technologies, Inc. Santa Clara, USA).

170 **2.6 Statistical analyses**

171 All data collected were subjected to analysis of variance (ANOVA) using SPSS Statistics for

172 Windows v17.0 (SPSS Inc., Chicago, USA). The data were first tested for normality; Levene's
173 test was used to determine equality of variances among the treatment groups. Statistical
174 significance was determined when $p \leq 0.05$. Tukey's highest significant difference (HSD) was
175 used for mean separation when a treatment effect was significant; data presented here are means
176 of at least four replicates. One aim of this work was to examine inter-relationships between plant
177 growth and yield variables, root and shoot nutrient concentrations vis-à-vis the salinity
178 treatments. The number of variables, however, was large (>30), therefore principal component
179 analysis (PCA) was used to reduce the dimensionality of the data by extracting and summarising
180 most of the variance in the multivariate data into a few dimensions. The variables analysed here
181 had different units (mass, area, number, etc.), so the PCA analyses used a correlation matrix as
182 input.

183 **3.0 Results**

184 **3.1 Growing conditions**

185 The temperature in the glasshouse fluctuated within 15% of their set values during the course of
186 the study. There was a spike in temperature in mid-July that caused the humidity to deviate by up
187 to 25% from the set range of 30–50%, otherwise the humidity remained within 10% of the
188 desired range throughout the study period. The photosynthetically active radiation (PAR) within
189 the glasshouse ranged between 260 and 900 $\mu\text{mol m}^{-2}\text{s}^{-1}$ during daylight hours.

190 **3.2 Plant growth characteristics**

191 Responses of vegetative and reproductive growth traits to salinity are summarised in Table 1.
192 Leaf production, leaf area and height of tomato plants were reduced on the saline soil and by
193 saline irrigation. On the saline soil, tomato plants were 12% shorter, had 25% fewer leaves that

194 had 73% smaller total area, compared with those on the non-saline soil. Relative chlorophyll
195 concentration, flower numbers and flower abortion in the tomato were insensitive to soil salinity.
196 Irrigation water salinity significantly reduced leaf area, numbers of leaves and flowers and plant
197 height, but increased relative chlorophyll concentration and flower abortion in this crop.

198 All growth variables in okra were reduced on the saline soil and by saline irrigation, while flower
199 abortion increased in response to the salinity treatments (Table 1). In okra, flower abortion
200 increased under salinity treatments, and more so than in tomato. Of all the traits examined in
201 both species, leaf area was the most sensitive to salinity irrespective of its source. Only weak
202 interactions were observed between soil and water salinity in their effects on the measured
203 variables in both crops, but were strong on chlorophyll concentrations in tomato.

204 **3.3 Fruit yield and quality**

205 Saline irrigation severely reduced the yield and yield components of tomato (Table 2). When
206 compared with the control, the 2.4 dS/m water salinity, reduced fruit yield by 88%, fruit number
207 by 77% and fruit size by 54%. Soil salinity also negatively affected tomato yield and yield
208 components, except the average fruit size. Water and soil salinity, however, increased sugar
209 concentration in tomato fruits, and for plants on the non-saline soil, irrigation with saline water
210 increased fruit sugar concentration by up to 34%, whereas on the saline soil, the increase was
211 74% (Table 2).

212 The yield and yield components of okra were significantly reduced by water and soil salinity; the
213 exception was fruit size (Table 2). Irrespective of soil salinity, increasing water salinity from 0.0
214 to 2.4 dS/m reduced fruit yield and number by more than 50%, but fruit size was comparatively
215 less sensitive. Okra lost more fruits on saline soil (19%) than the tomato (7%). Total fruit weight

216 per plant was the most responsive yield variable to both water and soil salinity in okra as in the
217 tomato. The yield response to irrigation water salinity was driven primarily via fruit number
218 whereas the response to soil salinity was almost equally driven both yield components.

219 **3.4 Total biomass production and its partitioning**

220 The dry weight of tomato plants fell significantly with water salinity on both saline and non-
221 saline soils (Table3). The weights of the plant components (roots, shoots and fruits) followed
222 similar trend in their response to water salinity. On both soils, water salinity reduced root/shoot
223 and fruit/shoot (putative harvest index). In contrast to water salinity, soil salinity had no
224 significant effect on plant dry weight or on its partitioning in the tomato.

225 The severity of adverse impact of salinity on plant dry weight and its components (roots, shoots
226 and fruits) in okra increased with water salinity, especially on the saline soil. Water salinity also
227 reduced root/shoot ratio but fruit/shoot ratios were unaffected. Soil salinity affected okra total
228 biomass, its components and partitioning (Table 3).

229 **3.5. Water use and water-use efficiency**

230 Water used by tomato was reduced on saline soil and by salinity of the irrigation water (Table 4),
231 and more so with water salinity (~17%) than soil salinity (6%). While water-use efficiency
232 (WUE) or the amount of fresh fruits produced for tomato per unit volume of water was not
233 affected by soil salinity, it fell with each increase in the salinity of irrigation water. The
234 deterioration in WUE with increasing salinity of the irrigation water was more severe on saline
235 soil than on non-saline soil. There were significant correlations between either the water-use or
236 WUE with water salinity:

237 Water-use: $y = -3.31x + 45.35, \quad r^2 = 0.81, \quad n = 24$

1a

238 WUE: $y = -2.61x + 7.69$ $r^2 = 0.59$, $n = 24$ 1b

239 Water use for okra was reduced by water and soil salinity (Table 4). On the non-saline soil,
240 water-use was only reduced when water salinity was raised to 2.4 dS/m, but on the saline soil
241 water-use was reduced with every increase in water salinity. On average, okra used about 11 L of
242 water less when grown on saline soil than on non-saline soil. The WUE for okra fell with every
243 increase in water salinity on the non-saline soil, dropping by 52% at the highest water salinity
244 treatment, while it declined by 43% with saline irrigation on the saline soil. There was, however,
245 no significant difference between the two soils in their mean WUE. The water-use and WUE
246 were related with water salinity as:

247 Water-use: $y = -0.334x + 36.8$, $r^2 = 0.45$, $n = 23$ 2a

248 WUE: $y = -0.53x + 2.22$, $r^2 = 0.33$, $n = 23$ 2b

249 **3.6 Elemental uptake and distribution**

250 Soil salinity did not alter nutrient concentrations in tomato tissues, but in the okra it increased
251 concentrations of Na and P in the roots and fruits, in addition to S in the roots (Fig. 1).
252 Concentration of nutrients in the root of tomatoes was in the order $Na > Ca \approx Mg > K > S > P$,
253 while in the shoot the order was $Na > K > Ca > Mg > P \approx S$. Elemental concentrations in the fruit
254 was dominated by Na on both saline and non-saline soils.

255 Soil salinity significantly increasing concentrations of Na, P and S in the root, P in the shoot and
256 Na and P in fruit in okra; Na was the dominant nutrient in both root and fruit, while Ca and K
257 dominated in the shoot (Fig. 1). Concentrations of Na and K in the roots, and of Ca, K and Mg in
258 the shoots, were higher for okra than found in tomatoes. Saline irrigation increased
259 concentrations of Na in all the three tissues of the plant, in addition to those of S in the root and

260 fruit, and of K, P and S in the shoot, in the tomato (Fig. 2a – c). Saline irrigation reduced
261 concentration of Ca, but increased that of K, in the shoot. In okra, saline irrigation increased
262 concentrations of Na in all the plant parts, and reduced those of Ca and K in the shoots (Fig. 2d –
263 f). Shoot concentrations of Na in okra was not more than a third that found in the tomato, while
264 those of Ca, K and Mg in okra were twice those in the tomato. In both crops, soil and water
265 salinity generally increased shoot/root Na concentrations, more so in the tomato in which the
266 ratio was 0.84 – 3.06 in saline conditions compared with 0.06–0.38 in okra (Table 3).

267 **3.7 Relationships between ionic concentrations and plant growth and yield variables**

268 Inter-relationships between root and shoot mineral nutrient concentrations, plant growth and
269 yield variables for each species are displayed along the first two orthogonal dimensions from
270 PCA for the two crops (Fig. 3). For tomato, the inter-relationships between the nutritional status
271 and plant traits are shown along the first two PCA dimensions, which jointly extracted about
272 60% of the total variance (Fig 3a). The first dimension (40% of the variance) reflects impact of
273 water salinity and shows that there were positive associations among the shoot P, K, S, Na, Cu,
274 Zn, Mn, root Na concentrations, and floret abortion (all with moderate to high positive loadings),
275 and all these were negatively correlated with fruit yield, water-use, WUE, fruit number per plant
276 as well as shoot Ca level (all with high negative loadings). The second dimension (20% variance)
277 revealed the impact of soil salinity. It contrasted root nutrient status (positive loadings) with leaf
278 number and area, plant height and floret abortion (all with negative loadings) to show a generally
279 inverse association between the two sets of variables. The impacts of the three water salinity
280 levels were distinctly separated, with hardly any overlaps amongst the symbols, along the first
281 principal dimension (Fig. 3b). The influences of the soil salinity treatments were apparent along
282 the second dimension albeit less distinctly, with some overlaps between blue and red symbols,

283 than observed with water salinity treatments.

284 For okra, the first dimension extracted 45% of the total variance as a measure of the impact of
285 water salinity on tissue nutrient concentrations, yield and growth variables (Fig. 3c). This
286 dimension reveals a negative correlation between root and shoot Na and P status (high negative
287 loadings and closely associated), on the one hand, and the plant growth and yield variables as
288 well as shoot concentrations of Mn, Mg, Ca, S and K status (high positive loadings), on the
289 other. There was thus a dichotomous association amongst these variables. In one group were Na
290 and P either in root or shoot that had negative associations with WU, WUE, fruit yield and
291 growth variables (chlorophyll on the 26th, leaf number and area, fruit number, and plant height).
292 In the other group were shoot concentrations of Ca, Mg, Mn, S, and K and root concentration of
293 K, all which had positive associations with the physiological, growth and yield variables. The
294 second dimension of portraying impact of soil salinity accounted for about a further 17% of the
295 variance; this had high loadings on root concentrations of Ca, Cu, Mg, Mn and Zn (Fig. 3c). The
296 variation represented by the second dimension was however only weakly associated with the
297 plant growth and yield variables. Overall impacts of soil and water salinity are clearly displayed
298 in figure 3c. It shows that the control and high (2.4 dS/m) irrigation were well separated, with
299 those of medium salinity overlapping with the other two, along dimension 1; there were
300 significant overlaps in the responses between the two soil salinity levels, especially with saline
301 irrigation, along dimension 2.

302 As would be expected, there were also strong associations among the physiological, plant growth
303 and yield variables. For example, the amount of water used per plant was closely related with the
304 number of leaves per plant, leaf area and functional state as indicated by the late season
305 chlorophyll concentrations. Similarly, a tight clustering was evident among fruit number and

306 yield per plant, plant height and water use efficiency. Differential impacts of water- and soil-
307 salinity were further illustrated in terms of their relative contributions to total variance, e.g., in
308 yield and yield components for both crops (Fig. 4). Overall, not more than 5% of the variance in
309 fruit yield and the main yield components for tomatoes were due to soil salinity compared to 10–
310 28% in okra. In contrast, water salinity accounted for at least 50% of the variance in yield and
311 associated components in tomatoes, much higher than a maximum of 40% variance accounted
312 for in okra.

313 **4.0 Discussion**

314 Both crops were adversely impacted by salinity, but they differed in their relative sensitivity to
315 the source of salinity. Soil salinity was less injurious to tomato, which experienced a yield
316 reduction of just 13% compared with 48% in okra on the saline soil relative to non-saline soil
317 (Table 2). The two crops also differed in their attributes that were more sensitive to soil salinity.
318 Vegetative attributes (height, leaf number and area) were adversely affected, while the
319 physiological and reproductive attributes (chlorophyll contents and number of flowers produced
320 and their survival) remained unaffected in the tomato on saline soil. This was contrary to
321 reductions in all the three categories of plant attributes in okra grown on the saline soil (Table 1).
322 The two crops, however, were affected by water salinity with both crops experiencing significant
323 reductions in yield with every step increase in salinity on both soils. Regression analyses (data
324 not presented) using pooled data for all treatments showed that fruit yield in tomato fell by
325 almost 124 g/plant (36% of yield under non-saline conditions) with every unit increase in water
326 salinity. Every unit increase in water salinity reduced yield in okra relative to non-saline
327 irrigation by 17–31 g/plant with an average of 28% fall. Thus, tomato was more sensitive to
328 saline irrigation.

329 The tomato showed a large tolerance to shoot concentration of Na. An increase in shoot/root Na
330 to 1.05 caused a loss of only 14% in fruit yield, on saline soil (Table 3). It was likely that the Na
331 in the shoot was sequestered in the vacuoles and away from the cytoplasm of the leaf, where
332 most biochemical processes occur, consistent with *tissue tolerance* mechanism of salinity
333 (Munns et al., 2006). Saline irrigation, however, increased tissue concentrations of Na
334 throughout the tomato plant, with shoot concentration doubling with every step up in the water
335 salinity treatment (Fig. 2) and raising shoot/root Na to as high as 3.06 (Table 3). It was probable
336 that such a high Na load would have overwhelmed the vacuolar capacity to sequester Na which
337 must have then 'leaked' into the cytoplasm of the leaf to impair growth processes. This appeared
338 to have occurred in the current study when shoot/root Na concentration exceeded the mean value
339 of 0.8 found on non-saline soil. The tissue tolerance in tomato could be associated with its large
340 capacity for osmotic adjustment that maintained osmotic potential of the leaf constant above -1.0
341 MPa even with saline irrigation of up to 7.4 dS/m (Pasternak et al., 1986).

342 In contrast to tomato, okra was more sensitive to shoot Na and so minimised partitioning this
343 nutrient to the shoot. The shoot/root Na concentration in okra did not exceed 0.35 in plants on
344 saline soil irrigated with saline irrigation, which was desirable since even the low shoot/root Na
345 concentration of 0.16 with 1.2 dS/m irrigation on non-saline soil reduced fresh fruit yields by
346 36%. Minimising the transfer of Na to the shoot (mainly leaves) by the okra was consistent with
347 *salt exclusion* mechanism for tolerating saline conditions. In this crop the fruits become a Na
348 sink almost as large as the roots when the crop was exposed to saline environments (Fig. 1 and
349 2).

350 The other factor in salinity responses in both crops is the role of other cations in either being
351 detrimental to yield or buffering the phytotoxic effects of Na. For instance, P concentration in

352 either shoot or roots was negatively, while Ca and Mg were positively correlated with fruit yields
353 and several other yield attributes in both crops (Fig. 3). Excessive tissue concentration of P in
354 okra was reported to induce deficiency of several micronutrients such as Zn and Mn (Loneragan
355 et al., 1981) that play key roles in enzyme systems and chlorophyll synthesis. Shoot P
356 concentration of 0.25% (2500 mg/kg) far exceeded the upper limits of 40 mg/kg found in several
357 studies (Akande et al., 2006).

358 Preferential accumulation of K over Na in the shoot (mostly leaves) is another mechanism
359 commonly associated with salinity tolerance in plants (Gorham et al., 1990). The biplots of our
360 data show the shoot concentration of K and yields for okra being on the same side of the
361 reference line on dimension one in the plot of vector loadings (Fig. 3). The shoot K/Na values
362 found here were much larger than K/Na values published for okra of not more than 2.0 even
363 under non-saline conditions (Saleem et al., 2011), possibly a result of high nutrient management
364 in the current study. Tissue K and yield and growth variables for the tomato were on the opposite
365 sides of the reference line on the first dimension suggesting an inverse relationship. It was
366 possible that K might have been antagonistic to uptake of other cations such as Ca and Mg in the
367 tomato since both ions had low shoot concentrations that were just fractions of those found in
368 okra (Fig. 2 and 3), or even when compared to 4% reported in several vegetable crops
369 (Maksimović and Ilin, 2012).

370 Increases in shoot Na in the two crops adversely affected growth and yield variables, including
371 developing flowers and fruits. Increased incident of flower abortion under saline conditions has
372 been widely reported for many plant species, including crops as varied as tomatoes (Ghanem et
373 al., 2009), chickpea (Krishnamurthy et al., 2011), sunflower (Francois, 1995) and jojoba
374 (Benzioni et al., 1992). The mechanism of flower abortion due to salinity is not fully understood,

375 but the results presented here reveal it could be the result of high concentrations of macro (K, P
376 and S) and micro-nutrients (Na, Cu and Zn) in the shoot of tomato (Fig. 3a).

377 Reductions in growth and associated processes due to salinity (Table 3), including water-use and
378 water-use efficiency (Table 4), are consistent with many previous studies on tomatoes
379 (Barbagallo et al. 2012) and okra (Adewoye et al., 2010; ul-Haq et al., 2012). Reductions in root
380 growth are often associated with low water and osmotic potential in the rhizosphere that then
381 impedes uptake of nutrient and water (Munns and Tester, 2008), thereby restricting root and
382 shoot growth that would have constrained water-use in both crops (Table 4). Soil and water
383 salinity both increased glucose content of tomato fruit as found in several earlier studies and was
384 associated with increased K concentration in the fruits (Machado et al., 2003; Yurtseven et al.,
385 2005) as we present here.

386 For both crops, the impact of soil salinity was much smaller than of saline irrigation, especially
387 for tomato. Under field conditions, preferential ion uptake from the less saline topsoil has been
388 invoked to explain differential plant growth responses to soil vs water salinity (Maas and
389 Hoffman, 1977). The extent to which such preferential water extraction explained the lower
390 phytotoxicity of the soil salinity in the current study is not clear since the roots proliferated the
391 whole of the 24 cm deep soil. Although it was possible that the frequent irrigation from the top
392 could have created a concentration gradient in the soil profile over time, it was more likely that
393 the dissolved salt in the irrigation water was more readily available since its addition coincided
394 with irrigation that increased water availability, which promoted absorption of dissolved salt by
395 the plant (Maksimović et al., 2010) in preference to the salt sourced from the soil.

396 These results suggest that contrasting irrigation strategies are needed to optimise productivity for
397 the two crops under saline conditions. The high sensitivity of tomato to irrigation salinity

398 suggests that reducing irrigation events, i.e. longer irrigation intervals, would minimise the
399 potential for the uptake and accumulation of salts dissolved in the irrigation water by plants. For
400 okra, frequent and regular over irrigation will leach out the salt and prevent its accumulation in
401 the root zone. Frequent irrigation with saline water of up to 4.9 dS/m, twice the maximum used
402 in the current study, maintained the matric potential in the root zone of silty clay above the
403 threshold of -30 kPa to maintain crop water-use (Wan et al., 2007).

404 **3 Summary and conclusions**

405 Tomato and okra differed in their responses to soil or water salinity. The tomato due to its
406 apparent inability to divert Na away from the shoot (mainly leaves), showed *tissue tolerance* in
407 maintaining reasonable yields even as shoot/root Na concentration rose to 0.8. This crop must
408 have sequestered the Na in the vacuoles of leaf tissues allowing maintenance of growth
409 processes, but the storage capacity of vacuoles would have been overwhelmed with increased
410 salt load due to water salinity. Okra was quite sensitive to shoot Na with yield significantly
411 reduced with shoot/root Na as low as 0.15. In okra, we found most tissue Na in fruits and little in
412 leaves, functioning as a *salt exclusion* mechanism. The yield penalty due to saline irrigation was
413 therefore more severe in the tomato that lost about 85% of its fresh fruits than in the okra that
414 lost an average of 64% of its fresh fruits. Saline irrigation was more injurious to plants than
415 water salinity in both crops, accounting for the overwhelming majority of variance, probably due
416 to greater availability to the plants of dissolved salt in irrigation water than in the soil.

417 These results suggest that contrasting irrigation strategies are needed to optimise productivity for
418 the two crops under saline conditions. The high sensitivity of tomato to irrigation salinity can be
419 managed by extending irrigation intervals to minimise opportunities for salt uptake and
420 accumulation. By contrast, frequent and regular over irrigation will leach out the salt and

421 prevents its accumulation in, the root zone to ensure high yields in okra.

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428

429 **References**

- 430 Adewoye, A.O., Okunade, D.A. and Adekalu, K.O. (2010). Assessing the yields and nutrient
431 uptake of okra (*Abelmoschus esculentus*) using diluted stabilised wastewater for irrigation
432 in south-western Nigeria. *Journal of Waste Water Treatment and Analysis*, 1: 104. doi:
433 10.4172/2157-7587.1000104.
- 434 Akande, M.O., Oluwatoyinbo, F.I., Kayode, C.O. and Olowokere, F.A. (2006). Response of
435 maize (*Zea mays*) and okra (*Abelmoschus esculentus*) intercrop relayed with cowpea (*vigna*
436 *unguiculata*) to different levels of cow dung amended phosphate rock. *World Journal of*
437 *Agricultural Science* 2: 119–122.
- 438 Ashraf, M. (2004). Some important physiological criteria for salt tolerance in plants. *Flora* 199:
439 361–376.
- 440 Barbagallo, R.N., Silvestro, D. and Patané, C. (2012). Yield, physicochemical traits, antioxidant
441 pattern, polyphenol oxidase activity and total visual quality of field-grown processing
442 tomato cv. Brigade as affected by water stress in Mediterranean climate. *Journal of the*
443 *Science of Food and Agriculture* 93: 1449–1457.
- 444 Benzioni, A, Nerd, A, Rosenärtner, Y and Mills D. (1992). Effect of NaCl Salinity on Growth
445 and Development of Jojoba Clones: I. Young Plants. *Journal of Plant Physiology* 139: 731–
446 736.
- 447 Coste, S, Baraloto, C, Leroy, C, Marcon, E, Renaud, A, Richardson, A.D., Roggy, J-C.,
448 Schimann, H., Uddling, J., Hérault, B. (2010). Assessing foliar chlorophyll contents with
449 the SPAD-502 chlorophyll meter: a calibration test with thirteen tree species of tropical
450 rainforest in French Guiana. *Annals of Forest Science* 67: 607 (5pp).

451 del Amor, F. M., Martinez, V. and Cerda, A. (2001). Salt tolerance of tomato plants as affected by
452 stage of plant development. HortScience 36: 1260 –1263.

453 Flowers, T. J. (2004). Improving crop salt tolerance. Journal of Experimental Botany 55: 307 –
454 319.

455 Francois, L (1995). Salinity effects on four sunflower hybrids. Agronomy Journal 88: 215–219.

456 Gorham, J, Bristol A, Young EM, Wyn Jones RG, Kashour G (1990). Salt tolerance in the
457 Triticeae: K/Na discrimination in barley. Journal of Experimental Botany 41: 1095–1101.

458 Ghanem, M.E., van Elteren, J., Albacete, A., Quinet, M., Martínez-Andújar, C., Kinet, J-M.,
459 Pérez-Alfocea, F., Lutts, S. (2009). Impact of salinity on early reproductive physiology of
460 tomato (*Solanum lycopersicum*) in relation to a heterogeneous distribution of toxic ions in
461 flower organs. Functional Plant Biology 36: 125–136

462 ul-Haq, I., Khan, A.A., Khan, I.A., Azmat, M.A. (2012). Comprehensive screening and selection
463 of okra (*Abelmoschus esculentus*) germplasm for salinity tolerance at theseedling stage and
464 during plant ontogeny. Journal of Zhejiang Univ-Sci B (Biomed & Biotechnol) 13: 533–
465 544

466 Krishnamurthy, L., Turner, N.C., Gaur, P.M., Upadhyaya, H.D., Varshney, R.K., Siddique,
467 K.H.M., Vadez, V. (2011). Consistent variation across soil types in salinity resistance of a
468 diverse range of chickpea (*Cicer arietinum* L.) genotypes. Journal of Agronomy and Crop
469 Science 197: 214–227.

470 Loneragan, J.F., Grunes, D.L., Welch, R.M., Aduayi, E.A., Tengah, A., Lazar, V.A., Cary, E.E.,
471 (1981). Phosphorus Accumulation and toxicity in leaves in relation to zinc supply. Soil
472 Science Society of America Journal 46: 345–352

- 473 Maas, E.V., Hoffman, G.J. (1977). Crop salt tolerance-current assessment. *Journal of Irrigation*
474 *and Drainage, Div. Civ. Eng.*, 103: 115–134.
- 475 Machado, R.M.A., Rosaria, M., Oliveira, G. and Portas, C.A.M. (2003). Tomato root
476 distribution, yield and fruit quality under subsurface drip irrigation. *Plant and Soil* 255:
477 333–341.
- 478 Maksimović, I. and Ilin, Ž. (2012). Effects of salinity on vegetable growth and nutrients uptake.
479 *Irrigation systems and practices in challenging environments*. < [http://www.intechopen.com/books/irrigation-systems-and-practices-in-challenging-environments/effects-of-](http://www.intechopen.com/books/irrigation-systems-and-practices-in-challenging-environments/effects-of-salinity-on-vegetable-growth-and-nutrients-uptake)
480 [salinity-on-vegetable-growth-and-nutrients-uptake](http://www.intechopen.com/books/irrigation-systems-and-practices-in-challenging-environments/effects-of-salinity-on-vegetable-growth-and-nutrients-uptake), 180. (accessed: December 18, 2013).
- 481
- 482 Maksimović, I., Putnik-Delić, M., Gani, I., Marić, J. and Ilin, Ž. (2010). Growth, ion
483 composition, and stomatal conductance of peas exposed to salinity. *Central European*
484 *Journal of Biology* 5: 682–691.
- 485 Munns, R., James, R.A., Läuchli, A. (2006). Approaches to increasing the salt tolerance of wheat
486 and other cereals. *Journal of Experimental Botany* 57: 1025–1043.
- 487 Munns, R., Tester, M. (2008). Mechanisms of salinity tolerance. *Annual Reviews of Plant*
488 *Biology* 59: 651–681.
- 489 Pasternak, D., De Malach, Y. and Borovic, I. (1986). Irrigation with brackish water under desert
490 conditions. VII. Effect of time of application of brackish water on production of processing
491 tomatoes (*Lycopersicon esculentum* Mill.). *Agricultural Water Management* 12: 149–158.
- 492 Perez-Alfocea, F., Albacete, A., Ghanem, M. and Dodd, I.C. (2010). Hormonal regulation of
493 source-sink relations to maintain crop productivity under salinity: a case study of root to
494 shoot signalling in tomato. *Functional Plant Biology* 37: 592–603.

- 495 Rogers, M.E., West, D.W. (1993). The effects of rootzone salinity and hypoxia on shoot and root
496 growth in trifolium species. *Annals of Botany* 72: 503–509
- 497 Saleem, A., Ashraf, M., Akram and N.A. (2011). Salt (NaCl)-induced modulation in some key
498 physio-biochemical attributes in okra (*Abelmoschus esculentus* L.). *Journal of Agronomy*
499 and Crop Science 197: 202–213.
- 500 Shannon, M. C., Grieve, C. M., Lesch, S. M. and Draper, J. H. (2000). Analysis of salt tolerance
501 in nine leafy vegetables irrigated with saline drainage water. *Journal of the American*
502 *Society for Horticultural Science* 125: 658–664.
- 503 Wan, S., Kang, Y., Wang, D., Liu, S. P. and Feng, L. P. (2007). Effect of drip irrigation with
504 saline water on tomato (*Lycopersicon esculentum* Mill) yield and water use in semi-humid
505 area. *Agricultural Water Management* 90: 63 –74.
- 506 Yunusa, I.A.M., Burchett, M.D., Manoharan, V., DeSilva, D.L., Eamus, D., Skilbeck, G.C.
507 (2009). Photosynthetic pigment concentration, gas exchange and vegetative growth of
508 selected monocots and dicots treated with contrasting coal fly ashes. *Journal of*
509 *Environmental Quality* 38: 1466 –1472.
- 510 Yurtseven, E. Kesmez, G.D. and Unlukara, A. (2005). The effects of water salinity and potassium
511 levels on yield, fruit quality and water consumption of native centre Anatolian tomato
512 species. *Agricultural Water Management* 78: 128 –135.

513 Table 1. Impact of soil and water salinity on plant growth variables for glasshouse grown tomatoes and okra.

Soil salinity (dS/m)	Water salinity (dS/m)	Leaf area/ plant (cm ²)	Leaves/ plant ¹	Final plant height (cm)	Chlorophyll content (µg/cm ²) at		Flowers/ Plant ²	Flower abortion (%)
					95 DAT	117 DAT		
<i>Tomato</i>								
0.0	0.0	406.6a	84.0a	145.8a	65.1a	31.9a	42.8a	79.3c
	1.2	350.0b	70.3b	137.0ab	67.1ab	38.7b	45.0ab	75.7bc
	2.4	272.5c	67.3bc	133.0bc	86.4c	47.3c	40.8c	93.8a
	<i>Mean</i>	<i>343.0A</i>	<i>73.8A</i>	<i>138.6A</i>	<i>72.3A</i>	<i>39.0A</i>	<i>42.8A</i>	<i>82.8A</i>
3.0	0.0	126.7a	59.8a	136.0a	65.7a	40.2a	41.0a	68.3c
	1.2	84.0b	59.3a	124.0b	60.3b	44.4ab	43.8ab	84.1b
	2.4	68.5c	48.5b	110.0c	60.9b	46.5b	40.5bc	92.9a
	<i>Mean</i>	<i>93.0B</i>	<i>55.8B</i>	<i>123.3B</i>	<i>62.4A</i>	<i>43.8A</i>	<i>41.7A</i>	<i>81.9A</i>
<i>Okra</i>								
0.0	0.0	108.7a	14.8 a	116.5a	68.2a	68.6a	11.8a	15.5c
	1.2	95.3b	11.0 b	101.8b	65.7ab	63.8b	6.8b	7.3bc
	2.4	73.1c	10.0 bc	76.8c	53.0c	52.7c	6.5b	22.8a
	<i>Mean</i>	<i>92.3A</i>	<i>11.9A</i>	<i>97.9A</i>	<i>62.0A</i>	<i>61.4A</i>	<i>8.3A</i>	<i>15.2A</i>
3.0	0.0	12.0a	14.5a	78.8a	51.4a	57.3a	8.3a	24.1c
	1.2	4.0b	7.0b	48.3b	54.9b	44.1b	9.3a	33.9b
	2.4	1.0c	5.3bc	31.3c	41.9c	42.5bc	6.0c	45.6ba
	<i>Mean</i>	<i>5.3B</i>	<i>8.9B</i>	<i>52.8B</i>	<i>49.2B</i>	<i>47.6B</i>	<i>7.8B</i>	<i>32.6B</i>

514 ¹ measured at 20 days after transplanting (DAT); for each crop, means in the same columns followed with the same letter(s) for a given soil salinity are statistically
 515 similar at $p \leq 0.05$; the lowercase letters compare means for water salinity levels, and uppercase letters compare means for soil salinity

516

517 Table 2. Impacts of soil and water salinities on fresh fruit yields and yield components for glasshouse grown tomato and okra.

Soil salinity (dS/m)	Water salinity (dS/m)	Fruits/plant	Total fruit yield/plant (g)	Weight/fruit (g)	Glucose content in tomato fruit (mmol/L)
<i>Tomato</i>					
0.0	0.0	9.0a	341.8a	37.7a	42.5a
	1.2	11.2ab	213.4b	19.7b	53.2b
	2.4	2.2c	49.9c	14.4c	56.9c
	<i>Mean</i>	7.4A	201.7A	23.9A	50.8A
3.0	0.0	11.7a	366.3a	30.9 a	50.7a
	1.2	6.5b	119.6b	18.1b	71.7b
	2.4	2.5c	38.1c	13.2c	88.0c
	<i>Mean</i>	6.9B	174.6B	20.7A	70.1B
<i>Okra</i>					
0.0	0.0	10.0a	107.9 a	11.5a	<i>nd</i>
	1.2	6.3b	69.8 b	10.7a	<i>nd</i>
	2.4	3.0c	45.5c	9.0b	<i>nd</i>
	<i>Mean</i>	6.4A	74.4A	10.4A	<i>nd</i>
3.0	0.0	6.3a	61.4a	9.1a	<i>nd</i>
	1.2	6.3a	27.3b	8.3b	<i>nd</i>
	2.4	3.0b	26.7c	8.9ab	<i>nd</i>
	<i>Mean</i>	5.2B	38.5B	8.7A	<i>nd</i>

518 *nd*, not determined; for each crop, means in the same columns followed with the same letter(s) for a given soil salinity are statistically similar at $p \leq 0.05$;
519 the lowercase letters compare means for water salinity levels, and uppercase letters compare means for soil salinity.

520 Table 3: Impacts of soil salinity and water salinity on the dry weights of plant tissues, and shoot/root ratio in Na concentrations for glasshouse grown
 521 tomato and okra.

Soil salinity (dS/m)	Water salinity (dS/m)	Root (g)	Shoot (g)	Fruit (g)	Total ¹ (g)	Root/shoot	Fruit/shoot	Shoot/root Na
<i>Tomatoes</i>								
0.0	0.0	12.9a	64.9a	1.3 a	79.1a	0.19 a	0.02 a	0.32b
	1.2	4.2b	62.9ab	1.1 ab	68.2b	0.06 b	0.017 ab	0.84a
	2.4	3.4bc	49.6c	0.7 bc	53.7c	0.06 b	0.014 bc	1.21a
	<i>Mean</i>	6.8A	59.1A	1.0A	67.0A	0.10A	0.017A	0.71B
3.0	0.0	4.9a	62.8a	1.3a	69.0a	0.07 a	0.020 a	1.05b
	1.2	3.2b	55.7ab	1.2b	60.1b	0.05 ab	0.021 b	1.28b
	2.4	2.6bc	39.8c	0.6bc	43.0c	0.06 bc	0.015 c	3.06a
	<i>Mean</i>	3.5A	52.7A	1.0A	57.3A	0.06A	0.018A	1.54A
<i>Okra</i>								
0.0	0.0	4.6a	17.4a	1.6a	23.6a	0.26a	0.09a	0.12a
	1.2	2.8b	12.5b	1.0b	16.3b	0.20b	0.08b	0.06b
	2.4	1.9c	8.6c	0.8c	11.3c	0.20b	0.09a	0.16a
	<i>Mean</i>	3.1A	12.8A	1.1A	17.0A	0.22A	0.08A	0.12B
3.0	0.0	3.1a	12.7a	1.3a	17.1a	0.20a	0.10a	0.20b
	1.2	0.7b	4.7b	0.8b	5.9b	0.10b	0.10a	0.20b
	2.4	0.5bc	4.2bc	0.5c	5.5c	0.10b	0.10a	0.38a
	<i>Mean</i>	1.4B	7.2B	0.8B	9.5B	0.13B	0.1B	0.24A

522 ¹sums of root, shoot and fruit at harvest; for each crop, means in the same columns followed with the same letter(s) at a given soil salinity are statistically similar at p ≤
 523 0.05; the lowercase letters compare means for water salinity levels, and uppercase letters compare means for soil salinity.

524 Table 4: Impacts of soil salinity and water salinity on water-use and water use efficiency for fruit yield
 525 (WUE) for glasshouse grown tomatoes and okra.

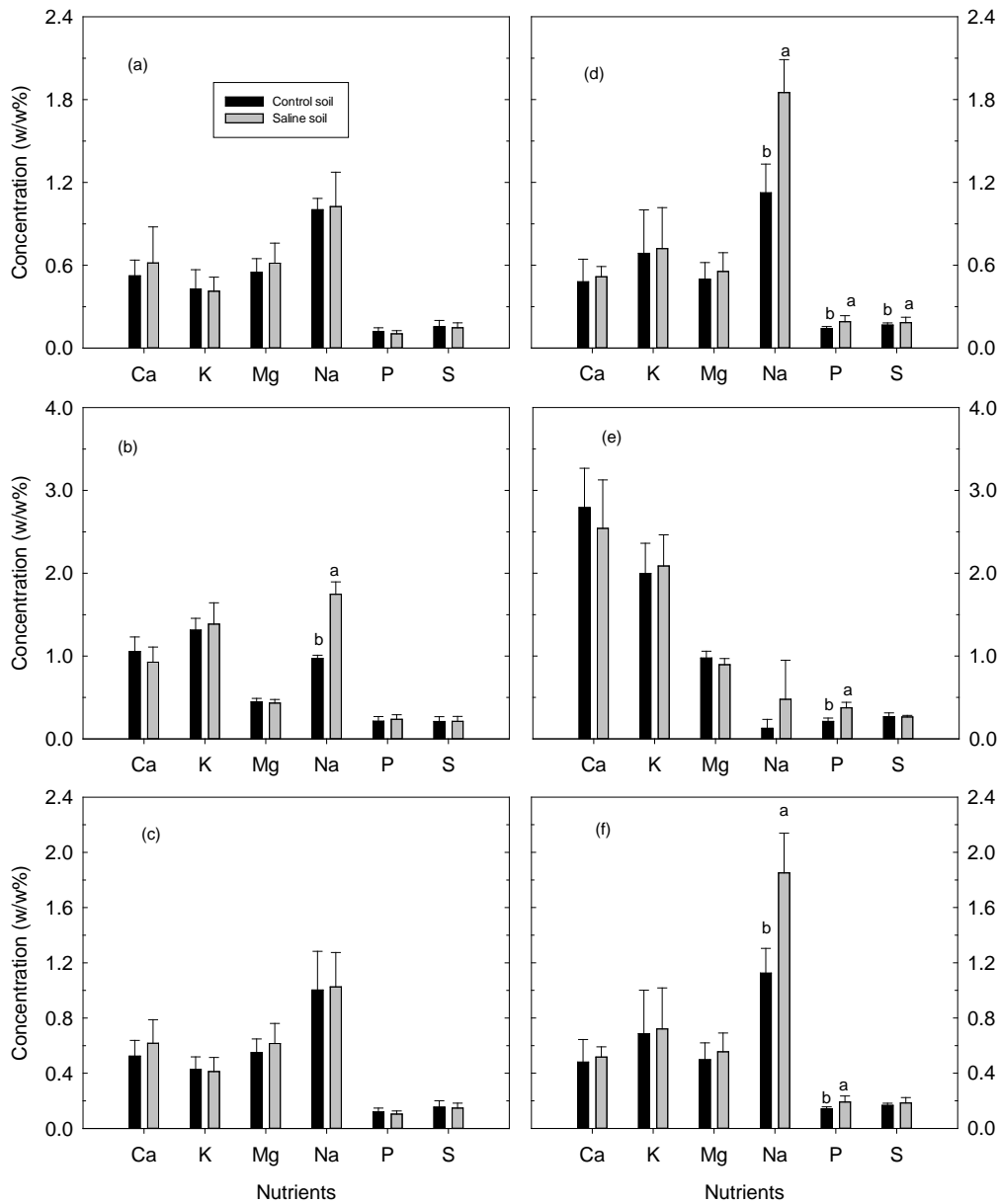
Soil salinity (dS/m)	Water salinity (dS/m)	Water use (L/plant)	W UE (g/L)
<i>Tomato</i>			
0.0	0.0	47.4a	7.1 a
	1.2	40.6b	4.5b
	2.4	38.1c	1.0c
	<i>Mean</i>	<i>43.3A</i>	<i>4.2A</i>
3.0	0.0	44.4a	7.6a
	1.2	39.9b	2.5b
	2.4	37.8c	0.8c
	<i>Mean</i>	<i>40.7B</i>	<i>3.6A</i>
<i>Okra</i>			
0.0	0.0	37.1a	2.9a
	1.2	36.6ab	1.9b
	2.4	32.2c	1.4c
	<i>Mean</i>	<i>35.3A</i>	<i>2.0A</i>
3.0	0.0	29.0a	2.1a
	1.2	25.0b	1.2b
	2.4	21.0c	1.2b
	<i>Mean</i>	<i>25.0B</i>	<i>1.5A</i>

526 For each crop, means in the same columns followed with the same letter(s) at a given soil salinity are statistically
 527 similar at $p \leq 0.05$; the lowercase letters compare means for water salinity levels, and uppercase letters compare
 528 means for soil salinity.

529

530

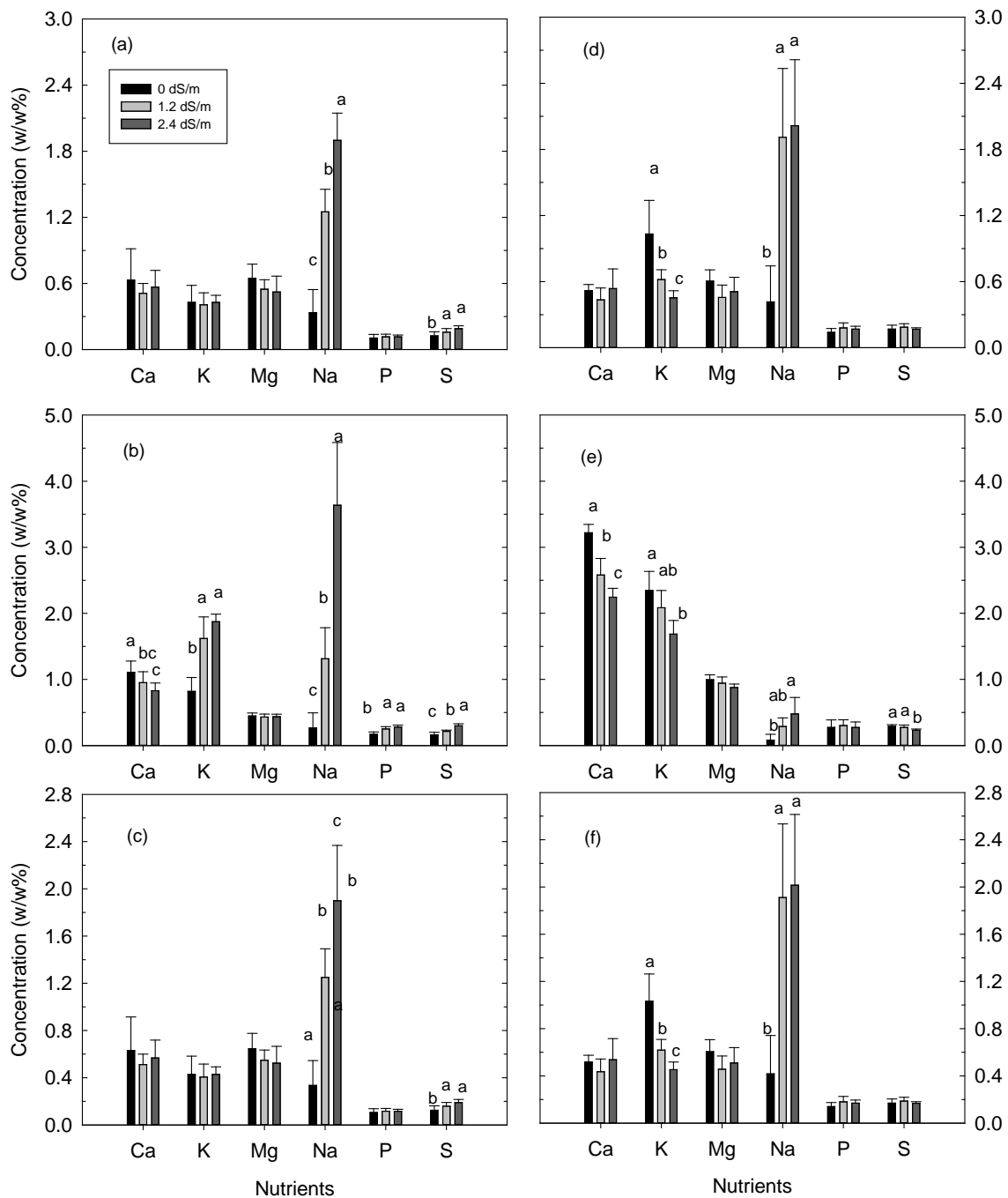
531



533

534 Figure 1. Impact of soil salinity on nutrient concentrations in the root (a, d), shoot (b, e) and fruit
 535 (c, f) at harvest for tomato (a – c) and okra (d – f). Where treatment means are significantly
 536 different ($p < 0.05$) are indicated by different letters.

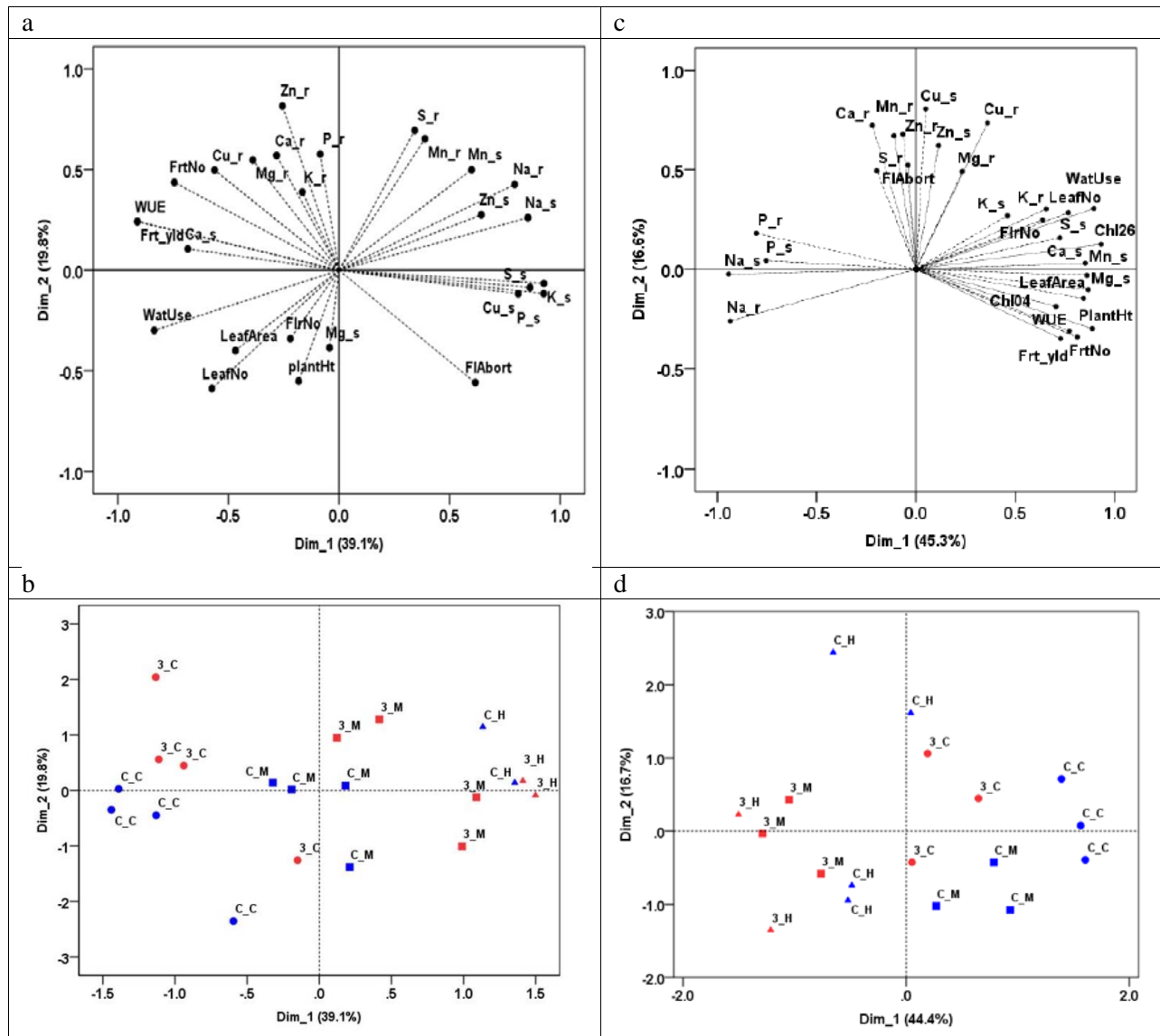
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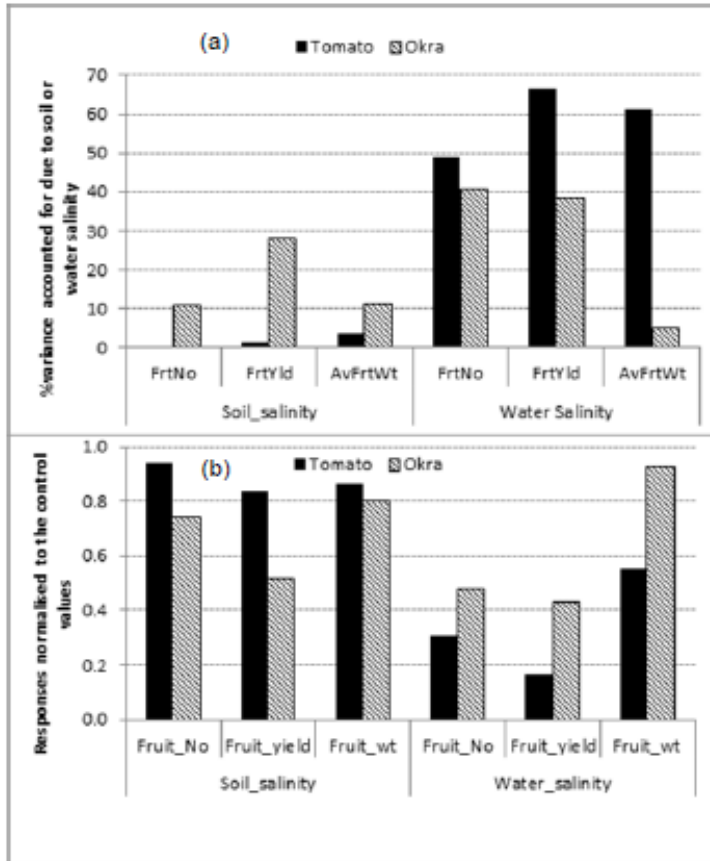
539 Figure 2. Impact of saline irrigation on nutrient concentrations in the root (a, d), shoot (b, e) and
 540 fruit (c, f) at harvest for tomato (a–c) and okra (d–f). Where treatment means are significantly
 541 different ($p < 0.05$) are indicated by different letters.

542



543

544 Figure 3. Interrelationships amongst plant response variables generated by principal component analyses (PCA)
 545 showing vector loadings (a, c) and biplots for salinity treatments (b, d) for tomatoes (a, b) and okra (bc, d). Codes in
 546 b and d are: C (control, 0 dS/m, circles) M (medium, 1.2 dS/m, squares) and H (high, 2.4 dS/m, triangles) irrigation
 547 water salinity, and C (control, 0 dS/m, blue) and 3 dS/m (red) soil salinity. The variables plotted are water-use
 548 (WU), leaf area (LA), leaf number (leafNo), chlorophyll concentrations (ch) on two dates, flower number (FlwrNo)
 549 and flower abortion (FlAbrt), plant height (PlantHt), fruit yield (FrYld) and fruit number (FrNo), and ionic
 550 concentrations in the shoot (_s) or root (_r).



551
 552 Fig. 4. Relative impacts of soil salinity and water salinity on selected yield variables for okra and
 553 tomato: (a) proportions of variance due to the respective salinity source, and (b) plant response
 554 variables normalized over control values.

555

556