Department of Chemical Engineering

PRETREATMENT OF WASTEWATER CONTAINING FATS AND OILS USING AN IMMOBILIZED ENZYME

HUANFEI JIA

"This thesis is presented as part of the requirements for the award of the Degree of Doctor of Philosophy of the Curtin University of Technology"

January 2002

DECLARATION

I certify that, this thesis contains no material which has been accepted for the award of any other degree or diploma in any university. To the best of my knowledge and belief this thesis contains no material previously published by any other person except where due acknowledgement has been made. This thesis is the result of my own work and contains nothing which is the outcome of work done in collaboration.

Huanfei JIA

28 January 2002

ABSTRACT

This thesis investigates an application of immobilized lipase for pre-treating wastewater containing fats and oils, which is difficult to treat practically. The kinetics of soluble lipase was studied for establishing background of the lipase. The immobilization of lipase was adopted in order to repeatedly use the expensive lipase. The developed immobilization methods were based on the characteristics of carriers, but covalent bonding of lipase was preferred because of strong adsorption nature. Three types of materials, nylon membrane and polystyrene-divinylbenzene and silica gel beads, were used for studying the lipase immobilization characteristics. The lipase from Canada rugosa was chosen because of its relatively high catalytic activity and commercial availability. The oily wastewater sources used were a simulated mixture of olive oil and distilled water as well as actual restaurant oily wastewater. A packed bed reactor packed with immobilized lipase was suitable for the study. Moreover, a comparative study of anaerobic digestion of lipase treated and un-treated oily wastewater was undertaken to evaluate the efficiency of the lipase pre-treatment method due to lack of the relevant literature in the enzymatic wastewater treatment field.

The kinetics of lipase catalyzed hydrolysis reactions was investigated in a stirred tank reactor. The experimental results confirmed that the lipase catalyzed reaction obeyed Michaelis-Menten model. The optimal pH and temperature of the lipase catalysed hydrolysis reaction were 7 and 37 °C, respectively. The conversion of oil to fatty acid was dependent on the reaction time and mass of the enzyme used. The lipase activities depended on the concentrations of some selected additives. Calcium ion improved lipase activity significantly amongst the additives used.

The immobilization of lipase was carried out using different materials, nylon membranes, polystyrene-divinylbenzene beads, and silica gel. Covalent adsorption was simple and successful for immobilizing the lipase onto nylon membrane which was pre-treated with HCl solution for releasing amino groups. The adsorption of lipase was completed after only a 2-hour reaction time. It was much more practical

for this shorter adsorption time (2 hours) rather than the 24 hours required for physical capillary adsorption of lipase.

The properties of the immobilized lipase and the performance of the reactors were compared amongst the soluble and immobilized lipase forms. The immobilization, particularly for covalent bonding, made lipase more resistant to thermal deactivation. It was evident that the optimum temperature was shifted from 37 °C for the soluble lipase to 45 and 40 °C for immobilized lipase adsorbed onto nylon and polystyrene-divinylbenzene beads, respectively. The immobilized lipase could be used repeatedly with only little activity loss. The repeatedly operational stability made the reuse of the immobilized lipase possible.

Comparison was also made between two types of beads, polystyrene-divinylbenzene beads and silica gels. Though polystyrene-divinylbenzene beads showed higher lipase activity and shorter adsorption time when compared to silica gels, the former beads were not suggested for large scale study because of high cost of the beads. One improvement achieved in this work was that the 24 hours required for silanization of silica gel was reduced to only a few hours using evaporating 3-APTES in acetone instead of refluxing 3-APTES in toluene.

It is worthwhile to point out that much higher enzyme activity was obtained using the packed bed reactor as against the membrane reactor when aqueous oil emulsion was fed into the reactors. The lipase activity was 64.2 % of soluble lipase activity for the immobilized lipase in the packed reactor but its activity was hardly detectable in the membrane reactor. Moreover, the operation of the packed bed reactor solved the oil separating problem that severely hampered the lipase catalytic activity in the membrane reactor in aqueous phase. This result suggests that the packed bed reactor with the immobilized lipase is applicable in treating oily wastewater.

The intrinsic parameters, V_{max} and K_m , were evaluated to study the internal diffusional effects of the porous spherical silica gel on the immobilized lipase. The changes of V_{max} and K_m for the immobilized lipase from those of the soluble lipase indicated that some alteration in the lipase intrinsic properties was caused by the immobilization of lipase. However, the magnitude of Thiele modulus suggested that the immobilized lipase was most likely reaction controlling. In addition, good agreement for V_{max} and K_m from experiments and numerical model estimations

seemed to suggest that the numerical model could be used for estimating V_{max} and K_m for the immobilized lipase.

An application was tried for conducting the hydrolysis of oily restaurant wastewater by soluble and the immobilized lipase. Enzyme activity of both forms was severely inhibited by the oily wastewater. The enzymatic activity was only 20 % and 15 % for soluble and the immobilized lipase, respectively, when compared to the initial activity value for the hydrolysis of olive oil by soluble lipase.

Evaluation of the efficiency for the proposed lipase pre-treatment method was carried out by monitoring the performance of two anaerobic digesters. These two digesters were fed with lipase treated and untreated restaurant wastewater that was neutralised with KOH solution prior to feeding. The oil-floating problem was minimised by this saponification of fatty acids with potassium hydroxide. However, there was no clear sign of an improvement for the treatment efficiency of the anaerobic digesters in terms of COD removal and methane production rate resulted in digesting lipase treated oily wastewater when compared to the one without lipase pre-treatment.

ACKNOWLEDGEMENTS

I would like to express my sincere gratitude to my supervisor, Associate Professor Ming Ang for his tremendous support, encouragement, and inspiring guidance throughout the course of my study. I gratefully thank Curtin University of Technology for the awards of Australian Postgraduate Scholarship.

I am also deeply grateful to the following people who have been a source of help to me during this study:

- Dr. P. Padmanabhan for her initial suggestions in the very early stage of my study.
- Mr. John Lauridsen, for his much appreciated technical assistance in the Chemical Engineering laboratory.
- All the technical workshop staff at the School of Engineering at Curtin University, for their work in the construction and maintenance of the laboratory equipment.
- Dr. Hasacawa Takahiro, from Shizuoka University in Japan for providing a numerical program and many useful discussions.
- The undergraduate students who have assisted in the experimental work.
- Mr.Budi Bisowarno, Ph.D student, for his very useful discussions and friendship in the course of my study.

Finally, I would like to thank my wife, Dr. Kaiyu Lin, for her understanding, great patience, and unconditional support. Without her company and the happiness she has brought me, this research would not have come to the end. My little angel daughter, Sophia, has been the greatest joy of my recent research life. Thanks are also given to two great people, my parents in law, who cooked delicious food which was always ready when I arrived home.

LIST OF PUBLICATIONS

- H.F. Jia, P. Padmanabhan and H.M. Ang. "Membrane Bioreactors-Potential Applications in wastewater treatment." Proceedings of CHEMECA 1998, Port Douglas, Australia, 149-154.
- H.F. Jia and H.M. Ang. "Hydrolysis of Olive Oil in a Batch Reactor: Additive Effects on Lipase Activity." Proceedings in APCCHE 1998, Seoul, South Korea, Vol 3, 1627-1630.
- H.F. Jia and H.M. Ang. "Performance of an Immobilised Lipase Membrane in Hydrolysis of Olive oil." Proceedings of CHEMECA 1999, Newcastle, Australia, 124-135.
- H.F. Jia and H.M. Ang. "Performance of an Immobilized Lipase onto Porous Beads in a Fixed Bed Reactor." Proceedings of CHEMECA 2000,636-672, Perth, Australia.
- H.F. Jia, H.M. Ang. "Immobilization of Lipase and Application of the Immobilized lipase" Submitted to Journal of Biotechnology, 2002.
- H.F. Jia, H.M. Ang. "Immobilization of Lipase and Estimation of the Kinetic Parameters for Immobilized Lipase." Submitted to Biochemical Engineering Journal, 2002.
- H.F. Jia and H.M. Ang. "Anaerobic Digestion of Pre-treated Lipid Rich Wastewater by Immobilized Enzyme". In preparation for submission to Water Science and Technology.

TABLE OF CONTENTS

DECL	ARATION	
ABSTI	RACT	
ACKN	OWLEDGEMENTS	
LIST (OF PUBLICATIONS	
TABL	E OF CONTENTS	
NOTA	TION	
LIST (OF TABLES	
LIST (OF FIGURES	
1 IN	TRODUCTION	1-1
1.1	BACKGROUND	1-1
1.2	Objectives	1-3
1.3	EXPERIMENTAL METHADOLOGY	1-4
1.4	ORGANISATION OF THE THESIS	1-5
2 LITI	ERATURE REVIEW	2-1
2.1	Introduction	2-1
2.2	EXISTING PROBLEMS IN TREATMENT OF FATTY AND OILY WASTEWATER	2-1
2.3	THE CURRENT METHODS FOR TREATING OILY WASTEWATERS	2-2
2. 3	2.1 Physical separation of the oils and fats	2-2
2. 3	2.2 Aerobic digestion of oily and fatty wastewater	2-3
2.3	Anaerobic digestion method for treating oils and fats	2-4
2.3	2.4 Enzymatic treatment of oily and fatty wastewater	2-6
2.4	GENERAL CHARACTERISTICS AND APPLICATIONS OF LIPASES	2-9
2.4	Advantages of enzymatic waste treatment	2-9
2.4	Characteristics and applications of lipases	2-9
2.5	IMMOBILIZATION OF LIPASES	2-12
2.5	I. Advantages of immobilized enzymes	2-12
2.5	5.2 Immobilization methods and characteristics of the immobilized lipase	s. 2-14
2.5	5.3 Characteristics of enzyme support materials	2-19

2.6 REVIEW OF IMMOBILIZED LIPASE REACTORS.......2-21

2.7 KINETICS OF HYDROLYSIS REACTIONS BY SOLUBLE AND IMMOR	BILIZED LIPASE 2-25
2.7.1 Hydrolysis reaction kinetics of the soluble lipase	2-25
2.7.2 Reaction kinetics of immobilized lipase	2-26
2.7.3 Dependence on chemical parameters	2-29
2.7.4 Dependence of physical parameters	2-30
2.8 CONCLUSIONS	2-31
3 HYDROLYSIS REACTION KINETICS CATALYSED BY	SOLUBLE
LIPASE	3-1
3.1 Introduction	3_1
3.2 MATERIALS AND METHODS	
3.2.1 Chemicals	
3.2.2 Equipment	
3.2.3 Analysis	
3.2.3.1 Expression of the enzyme activity	
3.2.3.2 Assay of the fatty acid	•
3.2.4 Experimental methods	
3.2.4.1 Hydrolysis of olive oil catalysed using lipase in batch	reactor3-7
3.2.4.2 Effect of additives on the lipase activity	3-7
3.3 ACCURACY OF THE ANAYLTICAL METHODS FOR FATTY ACID	3-7
3.4 SELECTION OF THE DESIRED LIPASE	3-8
3.5 THE ENZYME KINETICS MODEL	3-10
3.6 THE HYDROLYSIS OF OLIVE OIL USING SOLUBLE LIPASE	3-12
3.6.1 Determination of optimal pH and temperature	3-13
3.6.2 Effect of total mass loadings of lipase on reaction rate	3-14
3.6.3 Effect of the agitation speed on lipase reaction	3-14
3.6.4 Effect of the substrate concentration and water ratio on th	e reaction rate 3-15
3.6.5 Reaction progress for long-term hydrolysis reaction	3-16
3.7 ADDITIVE EFFECTS ON LIPASE CATALYTIC ACTIVITY	3-17
3.7.1 Effect of organic solvents on enzyme activity	3-18
3.7.2 Effect of surfactants on the lipase catalytic activity	3-19
3.7.3 The effect of inorganic salts on the lipase catalytic activity	,3-20
3.8 CONCLUSIONS	3-22

4.1	Introduction	4-1
4.2	MATERIALS AND EXPERIMENTAL METHODS	4-2
4.2	2.1 The materials	4-2
4.2	2.2 The equipment	4-3
4.2	2.3 Experimental methods for lipase adsorption onto membrane	4-3
4	1.2.3.1 Physical adsorption of lipase onto membrane strips	4-3
4	1.2.3.2 Immobilization of lipase onto membrane and on-line determ	nination of the
i	mmobilized lipase	4-4
4	4.2.3.3 The determination of fatty acids in hydrolysis reaction	4-5
1.3	DEVELOPMENT OF ANALYTICAL METHODS FOR DETERMINATION OF	IMMOBILIZED
LIPAS	SE	4-5
4.3	3.1 Modification of Lowery protein assay for determination of lipa	se4-6
4.3	3.2 On-line determination of lipase in bulk solution	4-8
4.4	SELECTION OF MEMBRANES	4-9
4.4	4.1 The preliminary adsorption of lipase onto membrane strips	4-10
4	4.4.1.1 Evaluation of the enzyme adsorption capacity in the membr	anes4-10
4	4.4.1.2 Evaluation of the hydrolysis activities of the immobilised en	nzyme4-11
1.5	EVALUATION OF THE IMMOBILISATION METHODS	4-13
4.5	5.1 The adsorption chemistry for nylons	4-14
4.5	5.2 The covalent immobilization procedure for nylon membranes	4-15
4.5	5.3 The adsorption characteristics	4-17
4	4.5.3.1 The adsorption time for both physical and chemical immobilistical and chemical and chemical immobilistical and chemical and chemi	ilization4-17
4	4.5.3.2 The adsorption capacity	4-18
4	4.5.3.3 The effect of the temperature on the immobilization	4-18
4.5	5.4 Effect of activated reagents and the adsorption stability	4-19
4.6	THE MEMBRANE REACTOR DESIGN	4-19
4.6	6.1 The configuration of the designed membrane reactor	4-21
4.7	THE PERFORMANCE OF THE IMMOBILIZED LIPASE IN THE MEMBRANE	
		4-21
17	7.1 The adsorption characteristics of the lipase onto nylon membra	

4.7.2 The performance of immobilised enzyme in biphasic organic and a	queous
media	4-23
4.7.2.1 The effect of flow rate on pressure drop	4-24
4.7.2.2 The effect of pH and temperature on catalytic activity	4-24
4.7.2.3 The catalytic activity of immobilised enzyme in hydrolysis of o	live oil
***************************************	4-25
4.7.2.4 The thermal and operational stabilities of the immobilised enzy	me4-26
4.7.3 The trials of immobilised lipase in the membrane reactor in aqueou	ı s
emulsion	4-27
4.8 CONCLUSIONS	4-28
5 IMMOBILISATION OF LIPASE ONTO POROUS BEADS	5-1
5.1 Introduction	5-1
5.2 Materials and methods	
5.2.1 The materials	
5.2.2 The equipment	
5.2.3 The adsorption methods	
5.2.3.1 The physical adsorption method	5-6
5.2.3.2 The chemical adsorption methods (for silica gels)	5-7
5.2.4 The hydrolysis of olive oil in the packed bed reactor	5-9
5.2.5 The analytical methods	5-10
5.3 THE SELECTION OF THE SUPPORT BEADS	5-10
5.3.1 Evaluation of the beads' adsorption capacity	5-10
5.3.2 Evaluation of the catalytic activities of the immobilised lipase	5-11
5.4 EXPERIMENTAL TRIAL OF A FLUIDIZED BED REACTOR	5-12
5.5 PERFORMANCE OF THE IMMOBILIZED LIPASE IN THE PACKED BED REACT	or5-12
5.5.1 The adsorption characteristics of the lipase onto the beads	5-12
5.5.2 The performance of immobilised enzyme in the packed reactor	5-13
5.5.2.1 The effect of flow rate on pressure drop and immobilized lipase	activity
	5-13
5.5.2.2 Effect of pH and temperature on lipase activity	5-13
5.5.2.3 The catalytic activity of immobilised enzyme	5-14

5.5.2.4 The thermal and operational stabilities of the immobilised	enzyme5-17
5.5.3 The regeneration of the porous beads	5-18
5.6 OPERATION OF A PACKED BED REACTOR WITH IMMOBILIZED LIPASE	ONTO SILICA
GEL	5-19
5.6.1 An alternative method for silanization of silica gel	5-19
5.6.1.1 Characteristics of the immobilized lipase onto activated sil	ica gels5-20
5.6.1.2 Performance of the immobilized lipase onto silica gels in the	ne packed bed
reactor	5-21
5.7 CONCLUSIONS	5-23
6 ESTIMATION OF INTRINSIC PARAMETERS FOR IMMOBI	LIZED
LIPASE IN POROUS BEADS	6-1
6.1 Introduction	6-1
6.2 Intrinsic parameters for soluble enzyme	
6.3 INTRINSIC KINETIC PARAMETERS FOR IMMOBILIZED ENZYME	
6.3.1 Theory for the immobilized enzyme system	
6.3.2 The calculation method for the numerical model	
6.4 RESULTS AND DISCUSSION	6-6
6.4.1 Michaelis-Menten kinetic parameters for the soluble lipase	6-6
6.4.2 Michaelis-Menten kinetic parameters for the immobilized lip	ase6-6
6.4.2.1 Approximate estimation of V_{max} and K_m from experiments	6-6
6.4.2.2 Estimation of V_{max} and K_m from the numerical model	6-8
6.5 CONLUSIONS	6-10
7 AN APPLICATION OF IMMOBILISED LIPASE FOR RESTAU	URANT
WASTEWATER TREATMENT	7-1
7.1 Introduction	7-1
7.2 MATERIALS AND EXPERIMENTAL METHODS	
7.2.1 The materials	
7.2.2 The equipment	
7.2.3 Experimental methods	
7.2.3.1 Hydrolysis of oils in the wastewater	
7.2.3.2 Start up of the batch anaerobic digesters	
7.2.4 Analytical methods	7-5

7.2.4.1 Analysis of fatty acids from oily wastewater hydrolysed by lipase	7-5
7.2.4.2 Assay the characteristics of the oily wastewater	7-5
7.3 THE CHARACTERISTICS OF THE RESTAURANT WASTEWATER	7-6
7.4 THE ENZYMATIC TREATMENT OF WASTEWATER USING SOLUBLE LIPASE	7-6
7.4.1 Hydrolysis of oily wastewater using soluble lipase	7-7
7.4.2 A trial for improvement of lipase activity	7-8
7.5 ENZYMATIC TREATMENT OF OILY WASTEWATER USING THE PACKED BED RE	ACTOR
	7-10
7.5.1 Hydrolysis of waste oils in oily wastewater	7-10
7.5.2 Improvement of the enzymatic activity of the immobilized lipase	7-10
7.5.2.1 Elevation of the reaction temperature	7-10
7.5.2.2 Addition of organic solvents	7-11
7.5.3 Extended period for the hydrolysis reaction	7-11
7.6 BATCH ANAEROBIC DIGESTION OF LIPASE-TREATED WASTEWATER	7-12
7.6.1 Anaerobic digester start up and operation	7-13
7.6.2 Evaluation of the improvement of the lipase pre-treatment efficiency	using
the anaerobic digesters	7-13
7.6.2.1 Apparent flotation problem of wastewater in the digesters	7-14
7.6.2.2 COD removal efficiency	7-14
7.6.2.3 Methane production	7-15
7.6.2.4 Oil removal efficiency	7-16
7.7 CONCLUSIONS	7-17
8 CONCLUSIONS AND RECOMMENDATIONS	8-1
8.1 Introduction	8-1
8.2 SIGNIFICANT FINDINFS DERIVED FROM THE THESIS	8-2
8.3 HYDROLYSIS REACTION KINETICS OF SOLUBLE LIPASE	8-3
8.4 CHARACTERISTICS OF THE IMMOBILIZATION METHODS	8-4
8.5 PROPERTIES OF THE IMMOBILIZED LIPASE AND THE PERFORMANCE OF THE	
REACTORS	8-4
8.6 EXTIMATION OF INTRINSIC PARAMETERS FOR STUDYIBG INTERNAL DIFFUSIO	
EFFECT IN POROUS BEADS	8-5

8.7	THE HYDROLYSIS OF OILS IN RESTAURANT WASTEWATER BY IMMOBILIZED LIP	ASE
		.8-6
8.8	EVALUATION OF THE PROPOSED LIPASE PRE-TREATMENT METHOD USING TWO	
ANAI	EROBIC DIGESTERS	.8-6
8.9	RECOMMENDATIONS FOR FURTHER RESEARCH	.8-6
FIGUI	RES	
REFE	RENCES	
APPE	NDIX	

NOTATION

B_I Biot number

BOD biological oxygen demand (mg/l)

COD chemical oxygen demand (mg/l)

D_e effective diffusivity (m²s⁻¹)

E free enzyme

ES enzyme and substrate complex

G geometric factor; 1 for slab, 2 for cylinder and 3 for sphere

 K_m intrinsic Michaelis-Menten constant (kg m⁻³)

N number of internal collocation points

n number of data sets

R radius of cylinder or sphere (m)

r iteration number

S_o initial substrate concentrate in a support (kg m⁻³)

S_b substrate concentration in a support (kg m⁻³)

 V_{max} intrinsic maximum reaction rate (kg m⁻³s⁻¹)

V_o initial reaction rate (kg m⁻³s⁻¹)

V experimentally measured initial reaction rate (kg m⁻³s⁻¹)

 \hat{V} Theoretical calculated initial reaction rate (kg m⁻³s⁻¹)

VSS volatile suspended solid (mg/l)

y dimensionless substrate concentration in a porous support

z dimensionless coordinate from centre of a support

 β_b dimensionless substrate concentration in a bulk liquid, $=S_b/K_m$

η effectiveness factor

 $\phi \qquad \text{Thiele modulus,} = \frac{R}{3} \sqrt{V_{max}/D_e K_m}$

LIST OF TABLES

- Table 2-1. Preparation and characteristics of immobilized enzymes
- Table3-1. Accuracy comparison of analysis of fatty acid both by titration and colorimetry methods.
- Table 3-2. Characteristics of the microbial lipases produced in Japan
- Table 3-3. The effect of organic solvents on lipase catalytic activity
- Table 3-4. The effect of surfactants on lipase catalytic activity
- Table 3-5. The effect of inorganic salts on lipase catalytic activity
- Table 4-1. Preparation of solutions for protein determination using Lowry assay
- Table 4-2. The results for physical adsorption of the lipase onto membrane strips
- Table 4-3. Characteristics of membrane materials for the immobilized lipases comparison with current work
- Table 4-4. The characteristics of immobilized lipase with various initial enzyme concentrations
- Table 5-1. The results for adsorption capacities and enzyme activities of lipase onto selected beads
- Table 5-2. Comparison of lipase activity between aqueous and organic media for lipase adsorbed onto polystyrene-divinylbenzene beads and silica gel
- Table 5-3. The properties and enzyme activity for the immobilized lipase onto silica gels
- Table 6-1: Parameters sets and experimental data (S_o and V_o) for executing the numerical program.
- Table 6-2. Converging results for V_{max} and K_m from executing the numerical program.
- Table 7-1. Characteristics of the oily wastewater from restaurants
- Table 7-2. Enzyme activities for different lipase forms in hydrolysis of olive oil and waste oils
- Table 7-3. Effect of additives on lipase activity in hydrolysis of waste oils
- Table 7-4. Operating parameters for both the anaerobic digesters
- Table 7-5. Effluent COD and methane produced both for the anaerobic digesters
- Table 7-6. Treatment performance summary for both the anaerobic digesters
- Table 7-7: Oil removal efficiency for both the anaerobic digesters

LIST OF FIGURES

- Figure 2-1. Schematic representation of the reactions catalyzed by lipases
- Figure 3-1. Schematic diagram of the stirred tank reactor used for free lipase
- Figure 3-2. Standard curve for oleic acid in 5 ml isooctane
- Figure 3-3. Comparison of fatty acid produced from different enzyme sources
- Figure 3-4. Enzyme activity obtained as a function of changing pH
- Figure 3-5. Enzyme activity obtained as a function of changing temperatures
- Figure 3-6. Fatty acid produced as a function of mass of lipase loads
- Figure 3-7. Enzyme activity obtained as a function of mass of lipase loads
- Figure 3-8. The effect of agitator speed on the hydrolysis reaction rate
- Figure 3-9. Fatty acid produced as a function of substrate concentrations
- Figure 3-10. Hydrolysis reaction rate as a function of substrate concentration
- Figure 3-11. The effect of water content on the oil hydrolysis reaction rate
- Figure 3-12. The fatty acid produced through long term hydrolysis reaction period
- Figure 3-13. The hydrolysis reaction rate versus reaction time for long term reaction course
- Figure 3-14. Percentage of conversion of olive oil to fatty acid for the long term hydrolysis reactions
- Figure 4-1. The standard Lowry curve for determination of protein
- Figure 4-2. The standard Lowry curve for determination of lipase
- Figure 4-3. The standard curve for on-line determination of lipase in solution
- Figure 4-4. Adsorbed lipase resulted from different adsorption methods versus adsorption time. (Adsorption types: physical adsorption; Glutaraldehyde and Carbodiimide modified covalent bonding)
- Figure 4-5. The degree of adsorption of lipase onto nylon membrane strips
- Figure 4-6. The adsorption capacity of enzyme onto nylon membranes
- Figure 4-7. The effect of the temperature on the lipase adsorption capacity performed on the nylon strips
- Figure 4-8. The stability of the immobilized lipase onto nylon membrane strips
- Figure 4-9a. Schematic diagram for the membrane reactor set up
- Figure 4-9b. Membrane module dimension (d)

- Figure 4-10. Bulk enzyme concentration versus adsorption time in the membrane reactor
- Figure 4-11. Amount of enzyme adsorbed versus adsorption time in the membrane reactor
- Figure 4-12. Variation of the pressure drop with flow rate in the membrane reactor.
- Figure 4-13. Effect of pH on catalytic activities of soluble and immobilized lipase
- Figure 4-14. Effect of temperature on catalytic activities of soluble and immobilized lipase.
- Figure 4-15. Fatty acid produced with various olive oil fed by the same immobilized lipase in the membrane reactor
- Figure 4-16. The extent of conversion of olive oil to fatty acids by soluble and immobilized lipase.
- Figure 4-17. Thermal inactivation of soluble lipase.
- Figure 4-18. Thermal inactivation of immobilized lipase
- Figure 4-19. The operational stability of the immobilized lipase by its repeated use in the membrane reactor
- Figure 4-20. The effect of water content in aqueous-oil mixture on the immobilized lipase activity in the membrane reactor
- Figure 5-1. The schematic diagram of a packed bed reactor with immobilized lipase onto beads
- Figure 5-2. Bulk lipase concentration versus adsorption time in the packed bed reactor with polystyrene-divinylbenzene beads
- Figure 5-3. Amount of lipase adsorbed onto polystyrene-divinylbenzene beads versus adsorption time in the packed bed reactor
- Figure 5-4. Effect of flow rate on pressure drops and enzyme activities of immobilized lipase onto polystyrene-divinylbenzene beads
- Figure 5-5. Effect of pH on catalytic activities of soluble and immobilized lipase.
- Figure 5-6. Effect of temperature on catalytic activities of soluble and immobilized lipase.
- Figure 5-7. Effect of organic solvent on catalytic activity of immobilized lipase
- Figure 5-8. Effect of substrate concentrations on fatty acid produced by the immobilized lipase in the packed bed reactor
- Figure 5-9. The comparison of fatty acid produced by immobilized lipase both in packed bed and membrane reactor in aqueous media

- Figure 5-10. The extent of conversion of olive oil to fatty acids by soluble and immobilized lipase
- Figure 5-11. Comparison of stability of immobilized lipase in repeated use of the lipase for the hydrolysis reactions conducted in the membrane and packed bed reactors
- Figure 5-12. Thermal inactivation of the immobilized lipase in packed bed reactor.
- Figure 5-13. Bulk lipase concentrations versus adsorption time for adsorption and readsorption of lipase onto polystyrene-divinylbenzene beads
- Figure 5-14. Amount of lipase adsorbed onto polystyrene-divinylbenzene beads versus adsorption time for adsorption and re-adsorption
- Figure 5-15. Amount of lipase adsorbed onto silica gels versus adsorption time from different adsorption methods in the packed bed reactor
- Figure 5-16. Effect of different adsorption methods on fatty acids produced by immobilized lipase onto silica gels
- Figure 5-17. Fatty acid produced from the immobilized lipase with various olive oil concentrations
- Figure 5-18. Comparison of fatty acid production by immobilized lipase using different beads
- Figure 5-19. Effect of particle sizes on lipase activity in the hydrolysis of olive oil catalyzed by lipase immobilized onto silica gels in the packed bed reactor
- Figure 6-1. Lineweaver-Burk plot for determination of V_{max} and K_m for the soluble lipase
- Figure 6-2. Hydrolysis reaction rate versus substrate concentrations catalyzed by lipase immobilized onto two sets of silica gels in the packed bed reactor
- Figure 6-3. Eadia-Hofstee plots for determination of V_{max} and K_m for lipase immobilized onto two sets of silica gels in the packed bed reactor
- Figure 7-1. The schematic diagram for an anaerobic digester system set up
- Figure 7-2. The calibration curve for quantifying methane concentration using infrared Spectrophotometer
- Figure 7-3. The hydrolysis of waste oils in restaurant wastewater by soluble lipase
- Figure 7-4. Comparison of fatty acids produced from the hydrolysis of waste oils and olive oil by soluble lipase
- Figure 7-5. Effect of CaCl₂ on lipase activity in the hydrolysis of waste oils in wastewater
- Figure 7-6. Effect of CaCl₂ on lipase activity in long term hydrolysis of oily

wastewater

- Figure 7-7. Effect of temperatures on lipase activity in the hydrolysis of oils in wastewater by immobilized lipase
- Figure 7-8. Effect of organic solvents on lipase activity in the hydrolysis of oils in wastewater by immobilized lipase
- Figure 7-9. Comparison of fatty acids produced in the hydrolysis of oils in wastewater by both the soluble and immobilized lipase
- Figure 7-10. Comparison of lipase activity in the hydrolysis of oils in wastewater by the soluble and immobilized lipase
- Figure 7-11. Degree of conversion of waste oils to fatty acids by soluble and immobilized lipase