

1 **Coral Reefs - Report**

2 **Comparative phylogeography of reef fishes from the Gulf of Aden to the Arabian Sea**

3 **reveals two cryptic lineages**

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33 **Abstract**

34 The Arabian Sea is a heterogeneous region with high coral cover and warm stable conditions at  
35 the western end (Djibouti), in contrast to sparse coral cover, cooler temperatures, and upwelling  
36 at the eastern end (southern Oman). We tested for barriers to dispersal across this region  
37 (including the Gulf of Aden and Gulf of Oman), using mitochondrial DNA (mtDNA) surveys of  
38 11 reef fishes. Study species included seven taxa from six families with broad distributions  
39 across the Indo-Pacific and four species restricted to the Arabian Sea (and adjacent areas). Nine  
40 out of 11 species showed no significant genetic partitions, indicating connectivity between  
41 contrasting environments spread across 2,000 km. One butterflyfish (*Chaetodon melannotus*) and  
42 a snapper (*Lutjanus kasmira*) showed phylogenetic divergences of  $d = 0.008$  and  $0.048$ ,  
43 respectively, possibly indicating cryptic species within these broadly distributed taxa. These  
44 genetic partitions at the western periphery of the Indo-Pacific reflect similar partitions recently  
45 discovered at the eastern periphery of the Indo-Pacific (the Hawaiian and the Marquesan  
46 Archipelagos), indicating that these disjunctive habitats at the ends of the range may serve as  
47 evolutionary incubators for coral reef organisms.

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## 55 **Introduction**

56 Phylogeographic analyses provide a unique means to detect historical and ecological processes  
57 that may not be apparent from contemporary species distributions (Palumbi 1997; Avise 2000).  
58 Such studies can reveal a diversity of outcomes, even among closely related species with similar  
59 life histories and geographic ranges (Bird et al. 2007; Gaither et al. 2010; Barber et al. 2011;  
60 Carpenter et al. 2011; DiBattista et al. 2012; Fouquet et al. 2012). Conversely, diverse taxa can  
61 also show concordant genetic patterns across broad spatial scales (Toonen et al. 2011; Selkoe et  
62 al. 2014). Multi-taxon studies across regions characterised by spatially and historically variable  
63 environmental conditions (i.e. comparative phylogeography) often generate a better  
64 understanding of how historical processes, environmental gradients, and ecological traits affect  
65 population structure and genetic diversity.

66 Comparative phylogeography has been used in many terrestrial and freshwater environments  
67 for delimiting regional phylogeographic patterns (Avise 1992; Hewitt 2000; Soltis et al. 2006;  
68 Waters et al. 2007; Bowen et al. 2016). However, resource and logistical limitations have  
69 restricted multi-taxon studies to only a few coral reef habitats. The Hawaiian Archipelago is a  
70 nearly linear habitat array extending 2,500 km with no obvious physical barriers or strong  
71 oceanographic discontinuities that might lead to hierarchical genetic structuring. Here, isolation  
72 by distance (IBD) was predicted to explain population structure, but multi-species studies across  
73 35 taxa found diverse patterns of genetic structuring including panmixia, chaotic genetic  
74 heterogeneity, regional structuring, as well as IBD (Selkoe et al. 2014). Reef taxa in the Coral  
75 Triangle (centred on Indonesia, the Philippines, and New Guinea) show a similar lack of

76 congruence in patterns of genetic structure, which may in part be driven by the complex  
77 geological history of the region (Barber et al. 2011; Carpenter et al. 2011). These studies show  
78 that, like terrestrial and freshwater systems, geological history and species-specific traits may  
79 play a role in determining patterns of population structure and genetic diversity in the coral reef  
80 environment, although this hypothesis has not been formally tested.

81 A major difference between marine and terrestrial or freshwater species is that marine  
82 species generally have much larger geographic ranges, which often span vast areas devoid of  
83 suitable habitat. Most reef-associated species also have a dispersive pelagic larval stage that can  
84 potentially connect populations across these uninhabitable areas. Contrary to expectations, the  
85 duration of the pelagic larval phase seems to be a poor predictor of population structure or range  
86 sizes (Weersing and Toonen 2009; Selkoe and Toonen 2011; Luiz et al. 2013; Riginos et al.  
87 2011, 2014; Gaither et al. 2016). Instead, latitudinal range (as a proxy for temperature tolerance),  
88 adult size (Luiz et al. 2012), species abundance (Strona et al. 2012), as well as behavioural  
89 factors (e.g. nocturnal activity and a tendency to school) correlate with range size (Luiz et al.  
90 2012, 2013). The importance of these factors may also vary depending on the size of the  
91 geographic range. A recent study of coral reef fishes indicates that small range endemics are  
92 under a suite of selection pressures that differ from widely distributed taxa, indicating that  
93 adaptation to local environmental conditions may also restrict species ranges (Gaither et al.  
94 2015). Consequently, endemics may be expected to show greater genetic structure than  
95 widespread species (Tenggardjaja et al. 2016), especially across regions that span strong  
96 environmental gradients or have a turbulent geological history.

97 Here we use a multi-taxon approach to investigate phylogeographic patterns in the distinct  
98 reef biota of the greater Arabian Sea, a vastly understudied region with diverse coral  
99 communities. The Arabian Sea is at the western margin of the Indo-Pacific and contains a  
100 contact zone between the distinct faunas of the Indo-Polynesian and Western Indian Ocean  
101 provinces (Briggs and Bowen 2012; DiBattista et al. 2015). The Western Indian Ocean province  
102 is bordered in the north by endemic hotspots in the Red Sea and the Arabian Gulf (DiBattista et  
103 al. 2016a), and represents one of the most geologically and oceanographically volatile regions of  
104 the world's tropical oceans (Cowman and Bellwood 2013; DiBattista et al. 2016a, 2016b).

105 Currently, the dominant environmental feature of the Arabian Sea region is the Indian Ocean  
106 monsoon system. The upwelling induced by the southwest monsoon brings changes in  
107 temperature and productivity, especially on the southern coast of Oman (Fein and Stephens  
108 1987), and reversals of monsoon-driven currents prompt strong seasonal variation in temperature  
109 and salinity in the Gulf of Aden (Sofianos et al. 2002; Smeed 2004). Subsequently, dramatic  
110 changes in reef habitats occur over distances of less than 2,000 km, which are well within the  
111 dispersal capacity of most reef fishes (Lessios and Robertson 2006; Keith et al. 2011, 2015). At  
112 the western extreme in the Gulf of Aden, the coastal waters of Djibouti have a high and relatively  
113 stable temperature regime with notable coral cover (Wilkinson 2008), and at the eastern extreme,  
114 the coastline of Oman supports rocky reefs with sparse coral cover and seasonal upwelling  
115 driving changes in productivity (Currie et al. 1973; Savidge et al. 1990; Sheppard et al. 1992;  
116 Barber et al. 2001; McIlwain et al. 2011). Due to the unique geomorphology of the coastline and  
117 seabed (e.g. Hanish Sill in the Strait of Bab al Mandab and Strait of Hormuz constricting the  
118 shallow Arabian Gulf), historic changes in sea level have caused significant alterations in habitat

119 availability, oceanographic currents, and environmental conditions (references from DiBattista et  
120 al. 2016a, 2016b). These fluctuations have had profound effects on the evolution of marine  
121 organisms across the region (DiBattista et al. 2016a, 2016b). Specifically, these forces have  
122 generated population genetic structure in some, but not all, species across the boundary of the  
123 Red Sea into the Gulf of Aden, indicating that historical conditions may have influenced each  
124 species differently (DiBattista et al. 2013; Fernandez-Silva et al. 2015; Ahti et al. 2016; Coleman  
125 et al. 2016; Waldrop et al. 2016).

126 Here we employ a multi-taxon approach to determine if there are concordant patterns of  
127 genetic structure across this environmentally heterogeneous region. Our sampling design crosses  
128 two previously described barriers to dispersal: the upwelling region off Oman (see Priest et al.  
129 2016), and the Strait of Bab al Mandab, which separates the Red Sea from the adjacent Gulf of  
130 Aden (DiBattista et al. 2016a, 2016b). We focus on 11 reef fishes from six families, with four  
131 range-restricted and seven widespread taxa, and test the hypothesis that endemic species are  
132 more likely to exhibit genetic structure than widespread species in a region where environmental  
133 conditions vary strongly across space and time..

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## 136 **Materials and methods**

### 137 *Sample collections*

138 We collected tissue samples (fin clip or gill filaments) from 11 reef fish species at sites in the  
139 greater Arabian Sea region while scuba diving or snorkelling between 2012 and 2015 (Fig. 1,  
140 Table 1). Geographic coordinates and sample sizes are provided in Table 2. Cases of low (or nil)

141 samples for a species from a particular site reflect rarity or complete absence in those locations,  
142 presumably owing to the heterogeneous nature of habitat in this region. Tissues were preserved  
143 in a saturated salt-DMSO solution or 96% ethanol, and subsequently stored at -20 °C. Select fish  
144 specimens were vouchered at the California Academy of Sciences (CAS; ESM Table S1).

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#### 146 *Mitochondrial DNA sequencing*

147 Total genomic DNA was extracted using the “HotSHOT” protocol of Meeker et al. (2007).  
148 Fragments of the mitochondrial cytochrome *c* oxidase subunit I gene (*COI*) were amplified using  
149 the primers FishF2 and FishR2 (Ward et al. 2005). Polymerase chain reaction (PCR) mixes  
150 contained 7.5 µl of BioMix Red (Bioline Ltd., London, UK), 0.26 µM of each primer and 5 to 50  
151 ng template DNA in 15 µl total volume. PCR conditions included an initial denaturing step at 95  
152 °C for 3 min, 35 cycles of amplification (30 s of denaturing at 94 °C, 60 s of annealing at 50 °C,  
153 and 60 s of extension at 72 °C), with a final extension at 72 °C for 10 min. PCR products were  
154 visualised on 2% agarose gels and purified by incubating with exonuclease I and FastAP™  
155 Thermosensitive Alkaline Phosphatase (ExoFAP; USB, Cleveland, OH, USA) at 37 °C for 60  
156 min, followed by 85 °C for 15 min. DNA was sequenced in the forward direction (and reverse  
157 direction for questionable haplotypes,  $N = 5$ ) with fluorescently labelled dye terminators  
158 following manufacturer’s protocols (BigDye; Applied Biosystems Inc., Foster City, CA, USA)  
159 using an ABI 3130XL Genetic Analyzer (Applied Biosystems). The sequences were aligned,  
160 edited and trimmed to a uniform length using GENEIOUS PRO 5.6.7 (Drummond et al. 2009).  
161 Individual mtDNA sequences are deposited in GenBank (accession numbers: XX-XX); mtDNA

162 sequences for *Chaetodon melapterus* were available from a related study in the region  
163 (DiBattista et al. 2015).

164 ARLEQUIN 3.5.1.2 (Excoffier et al. 2005) was used to calculate haplotype ( $h$ ) and nucleotide  
165 diversity ( $\pi$ ), as well as to test for population structure. jModelTest 1.0.1 (Posada 2008) was used  
166 to select the best nucleotide substitution model using the Akaike information criterion ( $AIC$ ).  
167 Genetic differentiation among sampling sites was first estimated with analysis of molecular  
168 variance (AMOVA) based on pairwise comparisons of sample groups; deviations from null  
169 distributions were tested with non-parametric permutation procedures ( $N = 99,999$ ). Pairwise  $\Phi_{ST}$   
170 statistics were also calculated in ARLEQUIN, significance tested by permutation ( $N = 99,999$ ) and  
171  $P$ -values adjusted according to the modified false discovery rate (FDR) method (Narum 2006).  
172 For  $\Phi_{ST}$  calculations, samples were pooled as follows given their close proximity ( $< 65$  km), lack  
173 of genetic differentiation (data not shown) and low individual sample size: 1) Bay de Ghoubett,  
174 Moucha/Maskali, and Obock in Djibouti, 2) Mirbat and Salalah in Oman and 3) Barr Al  
175 Hickman and Masirah Island in Oman.

176 Evolutionary relationships among haplotypes were evaluated using median joining spanning  
177 networks (Bandelt et al. 1999) in PopART 1.7 (<http://popart.otago.ac.nz>). In two cases  
178 (*Chaetodon melannotus* and *Lutjanus kasmira*; see Fig. 2), sequences from outside the Arabian  
179 Sea (see DiBattista et al. 2013) were included to highlight the presence of cryptic species within  
180 this region, which was not the case for other surveyed species. Deviations from neutrality were  
181 assessed with Fu's  $F_S$  (Fu 1997) for each species using ARLEQUIN; significance was tested with  
182 99,999 permutations.

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## 184 **Results**

185 *COI* sequences from 11 species of reef fish sampled across the Gulf of Aden to Arabian Sea  
186 included 2 to 42 haplotypes. Haplotype ( $h$ ) and nucleotide ( $\pi$ ) diversity ranged from 0.037 to  
187 0.879 and 0.00006 to 0.02933 (Table 1), respectively, with significantly higher nucleotide  
188 diversity values in widespread versus range-restricted fishes (two sample t-test,  $t = 2.09$ ,  $df = 40$ ,  
189  $P = 0.043$ ) but no discernable geographical trends across the Arabian Sea (One-way ANOVA,  $F$   
190  $= 0.75$ ,  $df = 6$ ,  $P = 0.61$ ; see Fig. 3). Higher nucleotide diversity values in widespread versus  
191 range-restricted fishes remained significant after excluding *L. kasmira* (two sample t-test,  $t =$   
192  $2.08$ ,  $df = 38$ ,  $P = 0.044$ ), an apparent outlier in the data set. This disparity in genetic diversity  
193 also does not appear to be driven by a sampling bias towards widespread versus range-restricted  
194 fish given a lower average sample size in the former (ESM Fig. S1). With two exceptions (*C.*  
195 *melannotus* and *L. kasmira*), the most common haplotype within a species was shared across  
196 sampling locations (Fig. 4). Analyses of molecular variance revealed significant population  
197 structure only for these two widespread species, *C. melannotus* ( $\Phi_{ST} = 0.813$ ,  $P < 0.001$ ) and *L.*  
198 *kasmira* ( $\Phi_{ST} = 0.838$ ,  $P < 0.001$ ), differentiated at Oman to Socotra and Djibouti to Somalia,  
199 respectively (also see ESM Table S2). These partitions are matched by divergences seen in the  
200 median joining spanning networks that include outgroup sequences (i.e. central Red Sea,  
201 Seychelles, and Maldives; see Fig. 2), invoking the possibility of cryptic species. The Red Sea to  
202 Djibouti lineage of *C. melannotus* is distinguished from a widespread Indian Ocean lineage  
203 (Socotra to Maldives) by five fixed substitutions ( $d = 0.008$ ). The divergence is much larger for  
204 *L. kasmira*, 22 fixed substitutions ( $d = 0.048$ ), distinguishing a Red Sea/Djibouti/Somalia to  
205 Seychelles lineage and a Socotra to Oman lineage, with both lineages observed in Djibouti.

206 Despite the strong genetic differences *within* both *C. melannotus* and *L. kasmira*, no  
207 morphological or meristic differences were observed in preliminary examinations by L.A.R (also  
208 see ESM Table S1). Tests for *COI* neutrality revealed negative and significant Fu's  $F_S$  values in  
209 7 of the 11 species (Fu's  $F_S = -27.39$  to  $4.29$ ; Table 1).

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## 211 **Discussion**

212 The study region, from the Gulf of Aden to the north-eastern coast of Oman, is characterised by  
213 environmental gradients and defined by historical barriers to gene flow (for review see DiBattista  
214 et al. 2016a, 2016b). To the west, the Strait of Bab al Mandab separates the endemism hotspot of  
215 the Red Sea from the adjacent Gulf of Aden. To the east, the coastline of Oman supports rocky  
216 reefs with sparse coral cover and is characterised by monsoonal upwelling that drives seasonal  
217 changes in productivity (McIlwain et al. 2011; DiBattista et al. 2016b). Mean sea temperatures  
218 on the Omani coast are significantly lower than the adjacent Gulf of Aden during the monsoon.  
219 The Yemeni island of Socotra, although influenced by the monsoon on its southern coast,  
220 supports carbonate reefs with notable live coral cover and a more stable annual sea temperature  
221 pattern on its northern coast (Kemp 1998, 2000). These environmental and geological factors are  
222 likely reflected in the genetic signatures of fish that inhabit these coral reefs.

223 Our mtDNA datasets reveal cryptic evolutionary divergences within two widespread reef  
224 fishes. In the butterflyfish *C. melannotus*, the mtDNA partition distinguished samples from the  
225 Red Sea and Djibouti by 0.8% at the *COI* barcoding gene compared to the widespread Indian  
226 Ocean lineage. In the snapper *L. kasmira*, a highly divergent lineage is apparent in the eastern  
227 Gulf of Aden and Oman (4.8% divergence from the widespread lineage). This lineage was not

228 detected in a previous range-wide survey of *L. kasmira* (Gaither et al. 2010), in which sampling  
229 in the Western Indian Ocean was limited to the Seychelles and South Africa. The Red  
230 Sea/Djibouti/Somalia lineage detected here is the same as the widespread lineage reported in  
231 Gaither et al. (2010). In the same study, Gaither and colleagues reported population genetic  
232 homogeneity across 12,000 km of the Pacific and Indian Oceans, therefore the genetic partition  
233 observed in the Arabian Sea is not due to limited dispersal ability. Interestingly, Gaither et al.  
234 (2010) described a cryptic evolutionary lineage on the eastern extreme of the range in the  
235 Marquesas Islands. This symmetry of divergent lineages at the eastern (Marquesas) and western  
236 (Socotra/Oman) ends of the range, and vast connectivity in between, adds to the accumulating  
237 evidence that peripheral habitats, especially those with unique environmental conditions, can  
238 serve as evolutionary incubators (Budd and Pandolfi 2010; Bowen et al. 2013; Hodge et al. 2014;  
239 Gaither et al. 2015).

240 The isolation of two lineages in the Arabian Sea is not surprising given the region's  
241 tumultuous paleo-climactic history and current heterogeneity in environmental conditions  
242 (DiBattista et al. 2016a, 2016b). Both Randall (1998) and Allen (2008) have noted that endemic  
243 hotspots for tropical marine organisms are located in peripheral areas of the Indo-Pacific. The  
244 Arabian Sea is bordered by two such hotspots (Red Sea and Arabian Gulf), which contain many  
245 species that have diverged from widespread sister taxa (Cowman and Bellwood 2011, 2013;  
246 Hodge and Bellwood 2016). This peripheral endemism has likely been augmented by historical  
247 sea level fluctuations and unique environmental conditions that may have isolated populations of  
248 widespread species (DiBattista et al. 2016a, 2016b), including *C. melannotus* and *L. kasmira*.  
249 The divergence of these cryptic lineages is estimated at 0.4 Ma and 2.4 Ma, respectively, which

250 date to the Pleistocene Epoch and thus the period of modern glacial cycles with frequent sea  
251 level fluctuations. In addition to historical isolation, the Arabian Sea is characterised by  
252 heterogeneous environmental conditions, also likely to have been a chronic condition during  
253 most of the Pleistocene (DiBattista et al. 2015). Thus, the genetic structure of *C. melannotus* and  
254 *L. kasmira* across the Arabian Sea may be explained by a combination of isolation and local  
255 adaptation, a hypothesis which warrants further investigation.

256 Congruence across species is seldom the case in comparative phylogeography studies of reef  
257 organisms (Lessios and Robertson 2006; Toonen et al. 2011; Selkoe et al. 2014). In the Coral  
258 Triangle, population genetic breaks and phylogenetic partitions appear in a variety of regions,  
259 with little discernible concordance (Barber et al. 2011; Carpenter et al. 2011; Sorenson et al.  
260 2014). Similar discordance was observed in the Hawaiian Islands, and analyses of various traits  
261 revealed that dispersal ability, taxonomy (fish versus invertebrates), and habitat specificity were  
262 significant predictor variables, although almost 90% of the variance remains unexplained (Selkoe  
263 et al. 2014). Closer to the Arabian Sea, DiBattista et al. (2013) showed genetic structure across  
264 the Strait of Bab al Mandab for some but not all species of reef fishes. For those species that did  
265 show structure, the depth of divergences showed no discernable concordance with each other or  
266 with glacial climate cycles. Although multi-taxon studies and comparative phylogeographic  
267 studies of reef organisms are few, some generalisations are emerging (Bowen et al. 2016). Those  
268 generalisations reinforced by this study include: 1) the geographic factors that define population  
269 structure in terrestrial and freshwater systems may not be as important in coral reef ecosystems,  
270 2) habitats at the periphery of the Indo-Pacific have higher endemism than previously expected,  
271 and 3) peripheral reef habitats like those contained in the Arabian Sea may host cryptic

272 evolutionary lineages, and thus genetic novelty, for even some of the most widely distributed  
273 species.

274 The most interesting finding of this study is that only two of the 11 species (*C. melannotus*  
275 and *L. kasmira*) had significant genetic structure and micro-evolutionary partitions across the  
276 Arabian Sea. Determining why this is the case will help elucidate the factors generating novel  
277 biodiversity in marine organisms. The two species that had genetic structure are widely  
278 distributed across the Indo-Pacific; however, five of the other nine species surveyed are also  
279 widespread and show no structure. A comparison of various biological and ecological traits  
280 thought to be associated with dispersal and colonisation success (Brown et al. 1996; Gaston  
281 2003; Luiz et al. 2012, 2013) such as geographic range size, body size, spawning mode, pelagic  
282 larval duration (PLD), diet, dietary specialisation, depth range, habitat use, schooling behaviour,  
283 and nocturnal activity did not reveal any obvious differences between species that did or did not  
284 display genetic structure (Table 3). Although *C. melannotus* is somewhat unique because its diet  
285 includes soft coral (Cole and Pratchett 2013), other specialist butterflyfishes (e.g. *C. melapterus*  
286 and *C. trifascialis*) did not show genetic structure. Furthermore, the only other species showing  
287 genetic structure (*L. kasmira*) is a generalist carnivore, like many other coral reef mesopredators  
288 (Hiatt and Strasburg 1960, Froese and Pauly 2016), including *C. argus* from this study, which  
289 itself did not show genetic structure. Contrasting patterns of genetic structure have also been  
290 reported in other closely-related, ecologically-similar, and widely-distributed reef fishes (e.g.  
291 Gaither et al. 2010) for reasons that remain unresolved.

292 Among the two cryptic evolutionary lineages revealed here, one lies primarily inside the Red  
293 Sea to the western Gulf of Aden and the other is distributed from the eastern Gulf of Aden to

294 Oman. Recent genetic surveys have found several more of these biodiversity gems hidden in Red  
295 Sea populations of widely-distributed species (DiBattista et al. 2013; Fernandez-Silva et al.  
296 2015, 2016; Coleman et al. 2016; Priest et al. 2016). These findings at the western periphery of  
297 the Indo-Pacific are remarkably concordant with surveys at the eastern periphery, which revealed  
298 cryptic evolutionary lineages at the Marquesas and Hawai'i (Gaither et al. 2010, 2011, 2015;  
299 DiBattista et al. 2011; Szabo et al. 2014; Bowen 2016). Together these data sets invoke a general  
300 finding that reef habitats in peripheral seas are important evolutionary incubators. As peripheral  
301 habitats lie at the geographic limits of a taxon's distribution, exposure to divergent  
302 environmental and habitat conditions are expected. Under such circumstances, strong and novel  
303 selection pressures are likely to contribute to genetic diversification (Gaither et al. 2011; Suzuki  
304 et al. 2016); the critical emerging issue is why this may involve only a proportion of the taxa  
305 present. The development of phylogeographic hypotheses based on taxon-specific traits (as per  
306 Papadopoulou and Knowles 2016) are therefore required to illuminate the diversification of  
307 evolutionary lineages in peripheral environments.

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**Table 1** Number of sample sites, fragment length and nucleotide substitution model (see Posada, 2008) for mitochondrial DNA cytochrome *c* oxidase subunit I (*COI*), Fu's  $F_S$  statistic, molecular diversity indices and differentiation metrics (i.e. AMOVA) for range-restricted and widespread reef fish sampled in the Arabian Sea region. All negative  $\Phi_{ST}$  values were adjusted to zero. Average values are  $\pm$  one standard deviation.

species	sites	fragment size (bp)	model	$N^b$	$H_N$	Fu's $F_S$	haplotype diversity ( $h \pm SD$ )	nucleotide diversity ( $\pi \pm SD$ )	$\Phi_{ST}$ (p-value)	evidence for barriers?
<u>range-restricted</u>										
<i>Chaetodon dialeucos</i> (Oman butterflyfish)	5	647	HKY	50	6	-2.81	0.505 $\pm$ 0.064	0.00089 $\pm$ 0.00081	0 (0.468)	No
<i>Chaetodon melapterus</i> (Arabian butterflyfish)	10	590	HKY	198	17	<b>-6.95<sup>a</sup></b>	0.723 $\pm$ 0.020	0.00261 $\pm$ 0.00174	0.008 (0.244)	No
<i>Chaetodon nigropunctatus</i> (black-spotted butterflyfish)	2	625	K80	54	2	-1.70	0.037 $\pm$ 0.035	0.00006 $\pm$ 0.00018	0.018 (0.404)	No
<i>Chaetodon pictus</i> (horseshoe butterflyfish)	10	582	TIM2	178	19	<b>-24.71</b>	0.404 $\pm$ 0.047	0.00089 $\pm$ 0.00083	0 (0.951)	No
<u>widespread</u>										
<i>Abudefduf vaigiensis</i> (Indo-Pacific sergeant)	9	576	K80	193	45	<b>-27.39</b>	0.879 $\pm$ 0.013	0.00341 $\pm$ 0.00215	0.013 (0.05)	No
<i>Cephalopholis argus</i> (peacock hind)	6	528	HKY	63	7	<b>-8.22</b>	0.211 $\pm$ 0.069	0.00042 $\pm$ 0.00056	0 (0.769)	No
<i>Chaetodon melannotus</i> (blackback butterflyfish)	4	619	K80	29	3	4.292	0.507 $\pm$ 0.079	0.00359 $\pm$ 0.00227	0.813 (< 0.001)	Yes (isolation at Socotra)
<i>Chaetodon trifascialis</i> (chevron butterflyfish)	8	577	TrN	117	12	<b>-5.74</b>	0.722 $\pm$ 0.028	0.00181 $\pm$ 0.00134	0.007 (0.333)	No

<i>Ctenochaetus striatus</i> (striated surgeonfish)	5	519	TrN	97	14	<b>-9.76</b>	0.624 ± 0.054	0.00183 ± 0.00140	0.011 (0.204)	No
<i>Halichoeres hortulanus</i> (checkerboard wrasse)	6	551	HKY	95	19	<b>-16.06</b>	0.611 ± 0.056	0.00211 ± 0.00152	0 (0.435)	No
<i>Lutjanus kasmira</i> (bluestripe snapper)	7	483	GTR	145	32	1.15	0.718 ± 0.034	0.02933 ± 0.01464	0.838 (< 0.001)	Yes (isolation at Djibouti & Somalia)

<sup>a</sup>Numbers in bold are significant,  $P < 0.02$  (Fu, 1997).

<sup>b</sup>Abbreviations are as follows:  $N$ , sample size;  $H_N$ , number of haplotypes.

**Table 2** Sample size and location for reef fish sampled in the Arabian Sea.

sampling site	location	<i>A.</i> <i>vaigiensis</i>	<i>Ce.</i> <i>argus</i>	<i>Ch.</i> <i>dialeucos</i>	<i>Ch.</i> <i>melannotus</i>	<i>Ch.</i> <i>melapterus</i>	<i>Ch.</i> <i>nigropunctatus</i>	<i>Ch.</i> <i>pictus</i>	<i>Ch.</i> <i>trifascialis</i>	<i>Ct.</i> <i>striatus</i>	<i>H.</i> <i>hortulanus</i>	<i>L.</i> <i>kasmira</i>
Moucha/Maskali, Djibouti	N 11.759° E 43.217°	20	20	–	1	20	–	21	20	19	20	25
Obock, Djibouti	N 11.967° E 43.333°	21	21	–	15	29	–	28	20	19	21	15
Bay de Ghoubett, Djibouti	N 11.533° E 42.667°	23	1	–	3	20	–	19	23	20	20	24
Berbera, Somalia	N 10.400° E 44.783°	16	13	–	–	16	–	20	14	15	12	21
Socotra, Yemen	N 12.617° E 54.350°	23	–	–	10	25	–	23	21	–	20	38
Salalah, Oman	N 16.912° E 53.960°	20	–	–	–	–	–	6	–	–	–	–
Mirbat, Oman	N 16.959° E 54.757°	–	1	7	–	12	–	16	7	–	–	–
Al Hallaniyats, Oman	N 17.483° E 55.983°	23	8	14	–	13	–	18	4	24	2	–
Schwaymeeyah, Oman	N 17.895° E 55.710°	–	–	2	–	–	–	–	–	–	–	–
Barr Al Hickman, Oman	N 20.383° E 58.217°	–	–	6	–	3	–	–	–	–	–	–
Masirah Island, Oman	N 20.165° E 58.634°	25	–	21	–	26	22	22	8	–	–	2
Muscat, Oman	N 23.525° E 58.740°	21	–	–	–	35	32	5	–	–	–	20

**Table 3** Biological and ecological traits associated with dispersal and colonisation abilities for the eleven study species. These include: geographic range size (extent of occurrence km<sup>2</sup>), body size (total length or TL) (Froese and Pauly 2016), spawning mode (Froese and Pauly 2016), pelagic larval duration (PLD) (Brothers and Thresher 1985; Victor 1986; Leis 1989; Thresher et al. 1989; Soeparno et al. 2012; Trip et al. 2014; J.P. Hobbs unpub. data) diet and dietary specialisation (Hiatt and Strasburg 1960; Bellwood and Pratchett 2013; Cole and Pratchett 2013; Froese and Pauly 2016), depth range (Froese and Pauly 2016), habitat use (Froese and Pauly 2016), schooling behavior, and nocturnal activity (Allen et al. 1998; Kuitert 2002; Yabutu and Berumen 2013).

species	Range <sup>a</sup> size (x 10 <sup>6</sup> km <sup>2</sup> )	Body size (TL in cm)	Spawning mode	PLD (mean days)	Diet	Dietary specialisation	Depth range (m)	Habitat use	Schooling behaviour	Nocturnal activity
<i>Chaetodon dialeucos</i> (Oman butterflyfish)	0.13	18	Broadcast	n/a	Omnivore (benthic invertebrates)	Generalist	5-25	Rocky and coral reefs	Pairs	No
<i>Chaetodon melapterus</i> (Arabian butterflyfish)	0.28	13	Broadcast	33.8	Corallivore	Moderate specialist	1-16	Coral reefs	Pairs	No
<i>Chaetodon nigropunctatus</i> (black-spotted butterflyfish)	0.11	14	Broadcast	n/a	Omnivore (benthic including coral)	Generalist	1-18	Rocky and coral reefs	Pairs	No
<i>Chaetodon pictus</i> (horseshoe butterflyfish)	0.24	20	Broadcast	39	Omnivore (benthic)	Generalist	1-20	Rocky reefs	Pairs	No
<i>Abudefduf vaigiensis</i> (Indo-Pacific sergeant)	50.2	20	Benthic	21.7	Omnivore (plankton and benthic)	Generalist	1-15	Rocky and coral reefs	Schools	No
<i>Cephalopholis argus</i> (peacock hind)	49.89	60	Broadcast	22.8	Carnivore (fish and benthic invertebrates)	Generalist	1-40	Coral reefs	Harems	Yes
<i>Chaetodon melannotus</i> (blackback butterflyfish)	49.02	18	Broadcast	n/a	Omnivore (benthic and soft corals)	Moderate specialist	1-20	Rocky and coral reefs	Solitary	No
<i>Chaetodon trifascialis</i> (chevron butterflyfish)	51.86	18	Broadcast	35.9	Corallivore	Extreme specialist	1-30	Coral reefs	Solitary	No
<i>Ctenochaetus striatus</i> (striated surgeonfish)	52.50	26	Broadcast	47 to 69	Detritivore	Generalist	1-34	Rocky and coral reefs	Schools	No

<i>Halichoeres hortulanus</i> (checkerboard wrasse)	50.75	27	Broadcast	32.5	Omnivore (benthic invertebrates)	Generalist	1-30	Coral reefs	Harems	No
<i>Lutjanus kasmira</i> (bluestripe snapper)	50.23	40	Broadcast	23.5	Carnivore (fishes and invertebrates)	Generalist	1-265	Coral Reefs	Schools	Yes

<sup>a</sup>Range size estimates were obtained through measurement of the area (km<sup>2</sup>) occupied by each species using IMAGE TOOL (as per Choat et al. 2012).

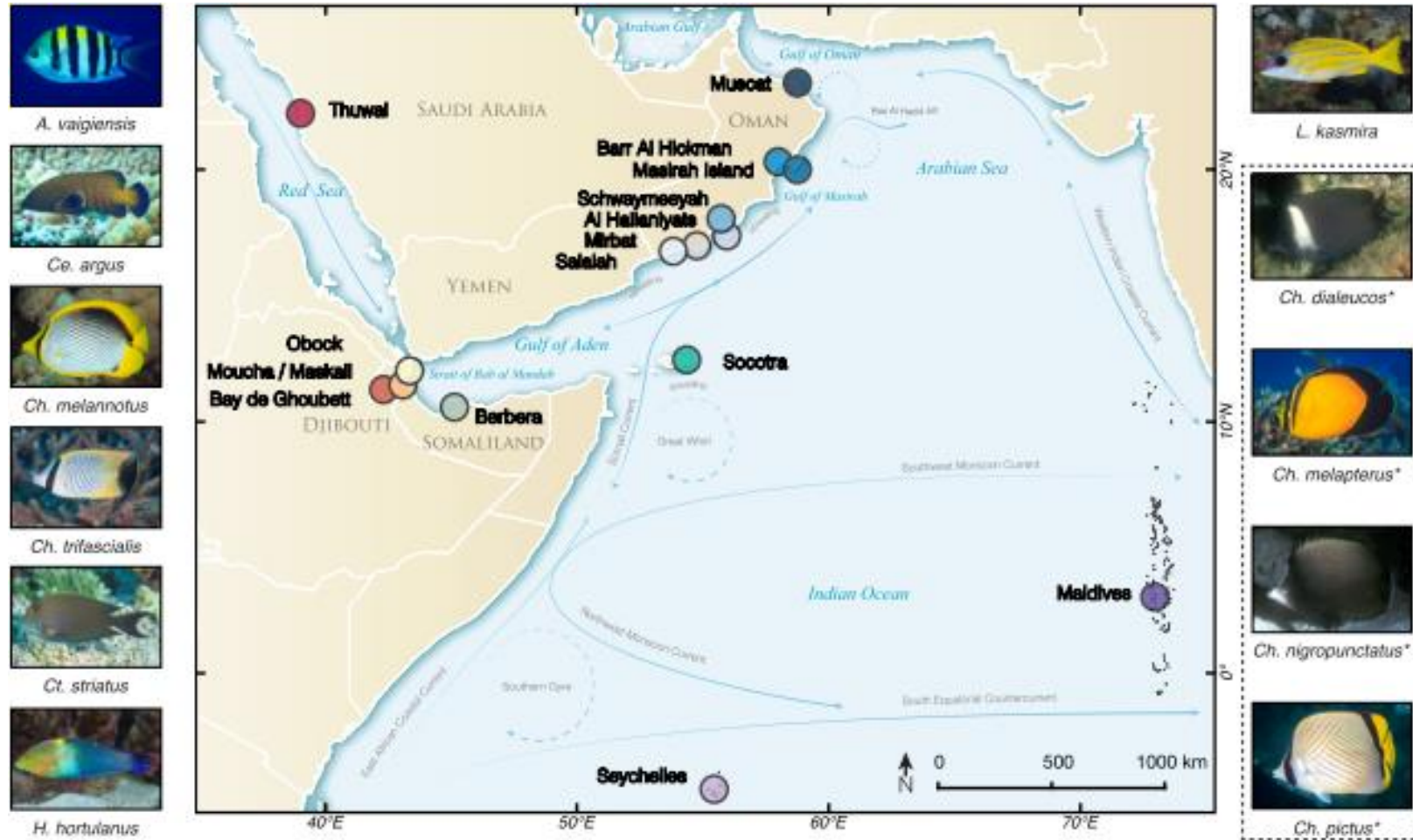
## Figures

**Fig. 1** Map indicating collection sites for all reef fish considered in this study, with range-restricted species denoted by asterisks. Colours used for collection location are identical to those in Fig. 2. Note the reversing circulation of the Somali Current (from northward to southward), the Southwest Monsoon Current (from westward to the eastward Northwest Monsoon Current), the Western Indian Coastal Current (from eastward to westward), and the current flowing into the Red Sea from the Gulf of Aden (versus out of the Red Sea and into the Gulf of Aden) during the northeast monsoon season (December to March). Site-specific samples sizes are provided in Table 2 (photo credit: T.H.S-T. and J.E. Randall). Samples from Thuwal, Maldives, and the Seychelles were included for only two species demonstrating cryptic lineages within Red Sea to Arabian Sea samples (*Chaetodon melannotus* and *Lutjanus kasmira*)

**Fig. 2** Median-joining networks showing relationships among mitochondrial DNA cytochrome *c* oxidase subunit I (*COI*) haplotypes for reef fish sampled in the Arabian Sea where cryptic evolutionary lineages were identified. *COI* fragment length and site-specific samples sizes are provided in Table 1 and Table 2, respectively. Outgroup populations are: *Chaetodon melannotus*, Thuwal, Kingdom of Saudi Arabia,  $N = 20$  and Maldives,  $N = 5$ ; *Lutjanus kasmira*, Thuwal, Kingdom of Saudi Arabia,  $N = 22$  and Seychelles,  $N = 20$ . Each circle represents a unique haplotype and its size is proportional to its total frequency (i.e. number of samples) as per the provided legend. Thin branches and black cross-bars represent a single nucleotide change, small open circles represent missing haplotypes and colours denote collection location as indicated by the embedded key

**Fig. 3** Haplotype (a) and nucleotide diversity (b) for 11 species of reef fish sampled from sites in the Gulf of Aden (Djibouti) to sites in the Arabian Sea (Muscat, Oman). Species and sample sizes are denoted by circle colours and sizes, respectively, as outlined in the provided key.

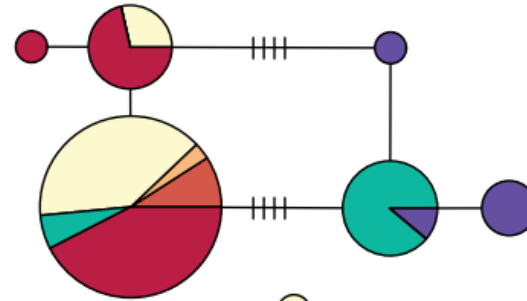
**Fig. 4** Median-joining networks showing relationships among mitochondrial DNA cytochrome *c* oxidase subunit I (*COI*) haplotypes for all remaining widespread and range-restricted reef fish sampled in the Arabian Sea region, with range-restricted species denoted by asterisks. *COI* fragment length and site-specific samples sizes are provided in Table 1 and Table 2, respectively. Each circle represents a unique haplotype and its size is proportional to its total frequency as per the included legend. Thin branches and black cross-bars represent a single nucleotide change, small open circles represent missing haplotypes and colours denote collection location as indicated by the embedded key.



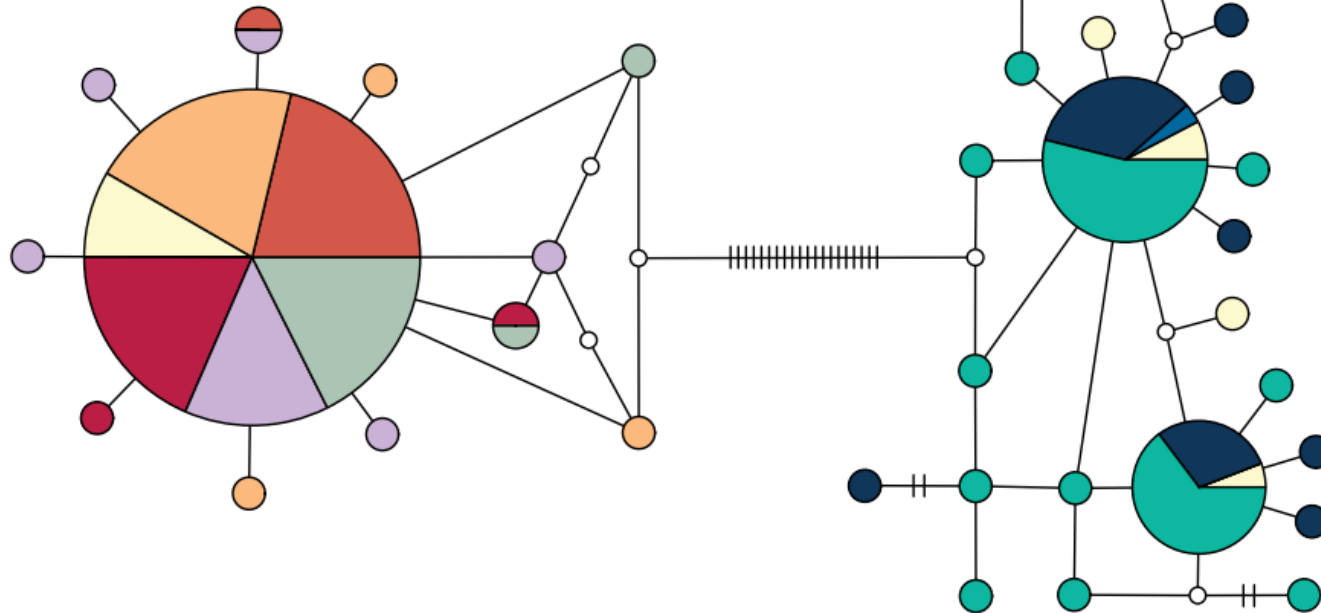


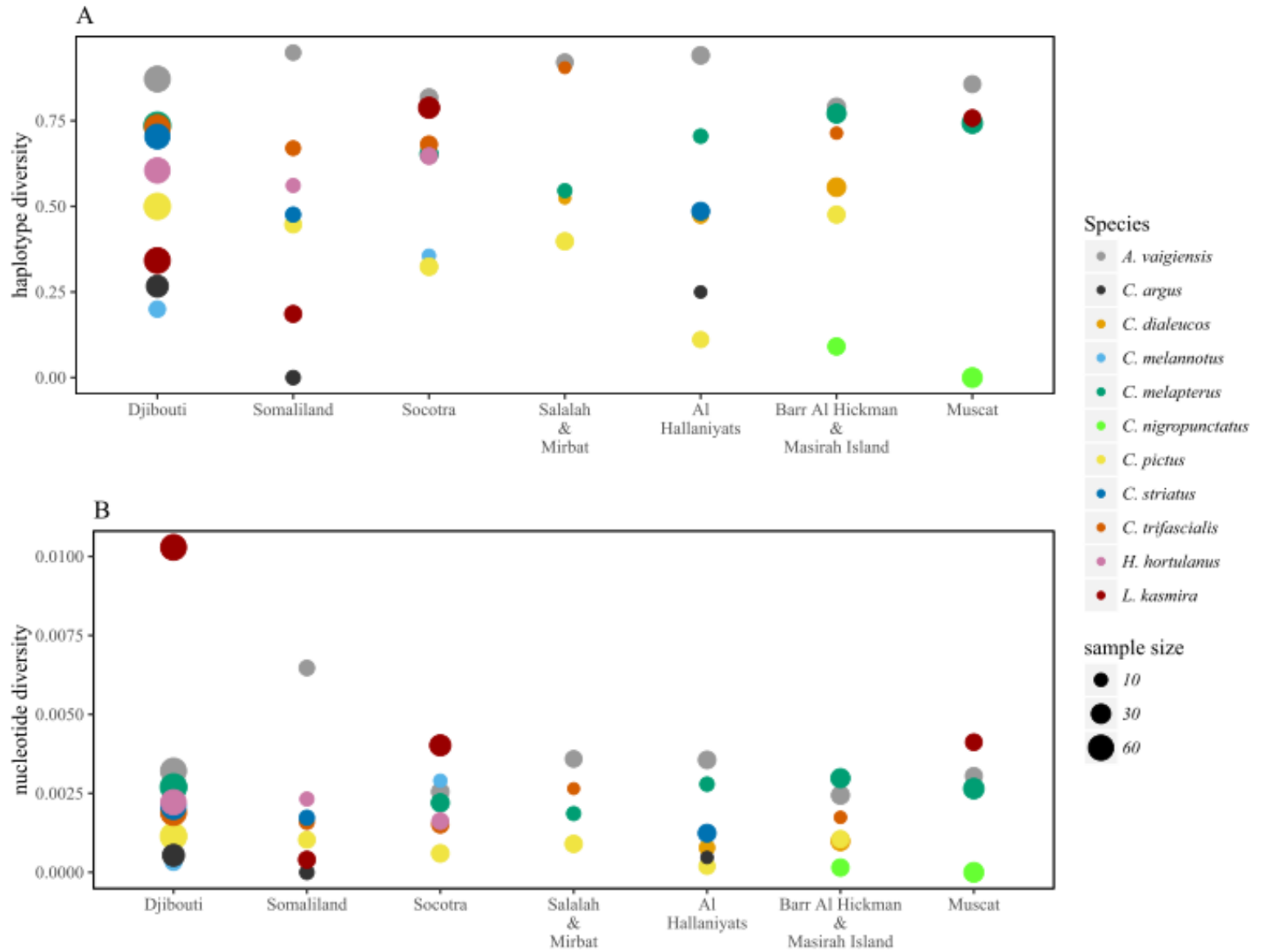


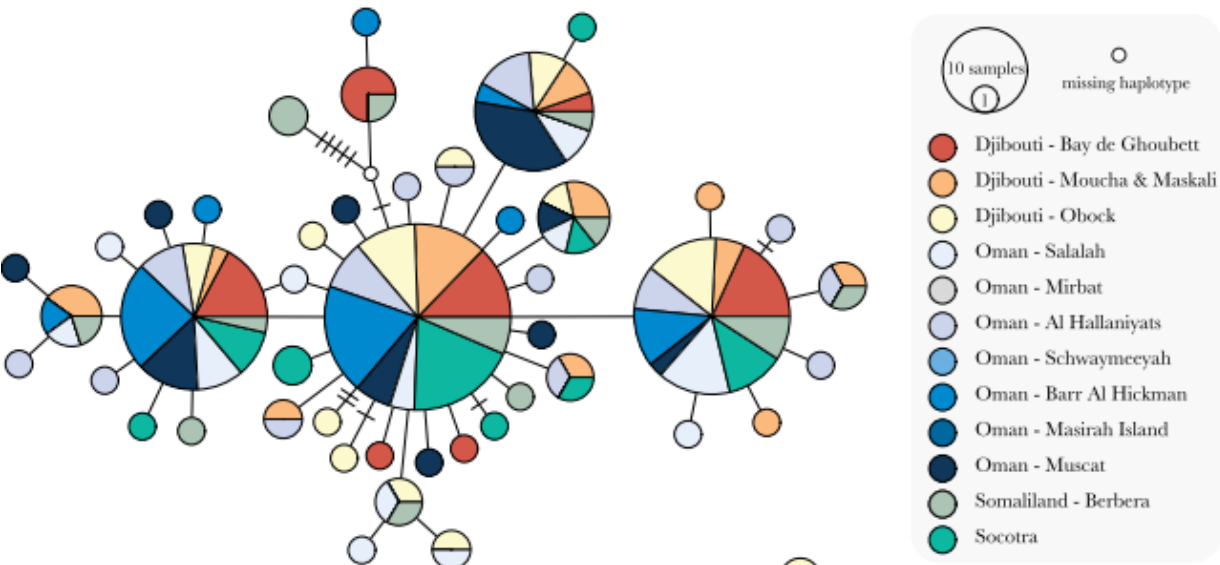
**A** *Chaetodon melannotus*



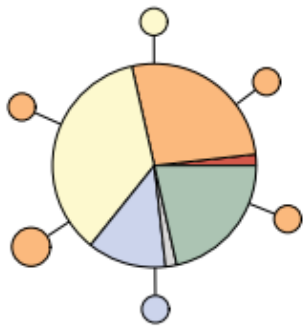
**B** *Lutjanus kasmira*



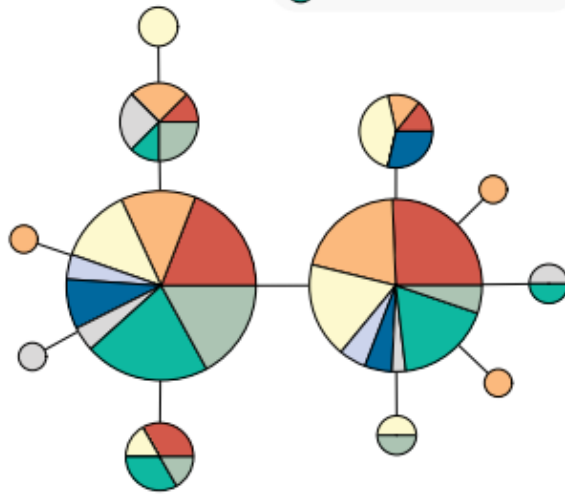




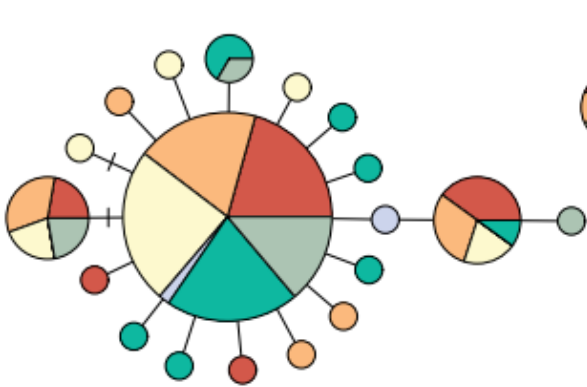
*Abudefduf vaigiensis*



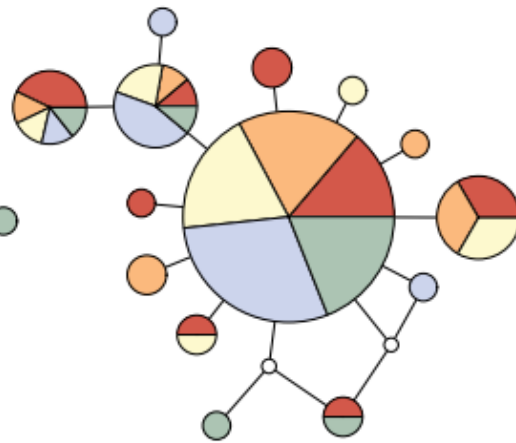
*Cephalopholis argus*



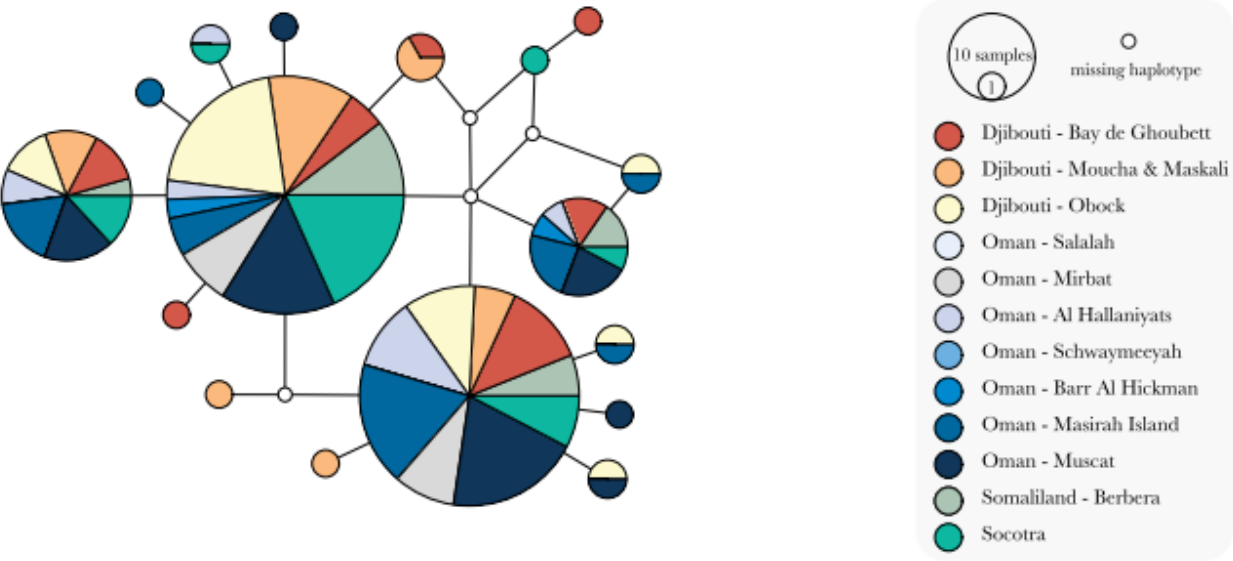
*Chaetodon trifascialis*



*Halichoeres hortulanus*



*Ctenochaetus striatus*

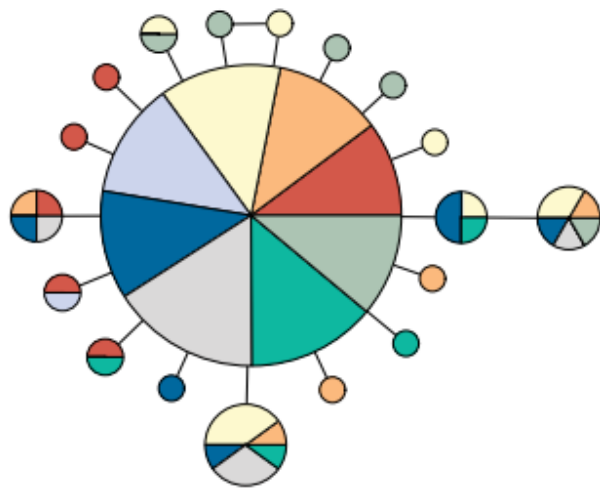


*Chaetodon melapterus*\*



*Chaetodon dialeucos*\*

*Chaetodon nigropunctatus*\*



*Chaetodon pictus*\*